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UNITED STATES PATENT APPLICATION

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FOR

SOLUBLE ZCYTOR11 CYTOKINE RECEPTORS

SOLUBLE ZCYTOR 11 CYTOKINE RECEPTORS

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REFERENCE TO RELATED APPLICATIONS

This application is related to Provisional Application 60/223,827, filed on August 8, 2000. This application is also related to Provisional Application 60/250,876, filed on December 1, 2000. Under 35 U.S.C. § 119(e)(1), this application
10 claims benefit of said Provisional Applications.

BACKGROUND OF THE INVENTION

Cytokines are soluble proteins that influence the growth and differentiation of many cell types. Their receptors are composed of one or more integral
15 membrane proteins that bind the cytokine with high affinity and transduce this binding event to the cell through the cytoplasmic portions of the certain receptor subunits. Cytokine receptors have been grouped into several classes on the basis of similarities in their extracellular ligand binding domains. For example, the receptor chains responsible for binding and/or transducing the effect of interferons (IFNs) are members of the type
20 II cytokine receptor family (CRF2), based upon a characteristic 200 residue extracellular domain. The demonstrated *in vivo* activities of these interferons illustrate the enormous clinical potential of, and need for, other cytokines, cytokine agonists, and cytokine antagonists.

25 DESCRIPTION OF THE INVENTION

The present invention fills this need by providing novel cytokine receptors and related compositions and methods. In particular, the present invention provides for an extracellular ligand-binding region of a mammalian Zcytor11 receptor, alternatively also containing either a transmembrane domain or both an intracellular
30 domain and a transmembrane domain.

Moreover, the present invention fills this need by providing novel soluble cytokine receptors that can be used to antagonize the effects of T-cell inducible factor (IL-TIF) in certain human disease states. In particular, the present invention provides for an extracellular ligand-binding region of a mammalian Zcytor11 receptor,
5 that is either homodimeric, heterodimeric, or multimeric.

Within one aspect, the present invention provides an isolated polynucleotide that encodes a soluble cytokine receptor polypeptide comprising a sequence of amino acid residues that is at least 90% identical to the amino acid sequence as shown in SEQ ID NO:3, and wherein the soluble cytokine receptor
10 polypeptide encoded by the polynucleotide sequence binds or antagonizes IL TIF (SEQ ID NO:8). In one embodiment, the polynucleotide is as disclosed above, wherein the soluble cytokine receptor polypeptide encoded by the polynucleotide forms a homodimeric receptor complex.

Within a second aspect, the present invention provides an isolated
15 polynucleotide that encodes a soluble cytokine receptor polypeptide comprising a sequence of amino acid residues that is at least 90% identical to the amino acid sequence as shown in SEQ ID NO:3, wherein the soluble cytokine receptor polypeptide encoded by the polynucleotide forms a heterodimeric or multimeric receptor complex. In one embodiment, the polynucleotide is as disclosed above, wherein the soluble
20 cytokine receptor polypeptide encoded by the polynucleotide forms a heterodimeric or multimeric receptor complex further comprising a soluble Class I or Class II cytokine receptor. In another embodiment, the polynucleotide is as disclosed above, wherein the soluble cytokine receptor polypeptide encoded by the polynucleotide forms a heterodimeric or multimeric receptor complex further comprising a soluble CRF2-4
25 receptor polypeptide (SEQ ID NO:33) or a soluble IL-10 receptor polypeptide (SEQ ID NO:34), or a soluble DIRS1 receptor polypeptide (SEQ ID NO:35). In another embodiment, the polynucleotide is as disclosed above, wherein the soluble cytokine receptor polypeptide encoded by the polynucleotide forms a heterodimeric or multimeric receptor complex further comprising a soluble CRF2-4 receptor polypeptide
30 (SEQ ID NO:33) or a soluble IL-10 receptor polypeptide (SEQ ID NO:34), or a soluble DIRS1 receptor polypeptide (SEQ ID NO:35).

Within a third aspect, the present invention provides an isolated polynucleotide that encodes a soluble cytokine receptor polypeptide comprising a sequence of amino acid residues as shown in SEQ ID NO:3, wherein the soluble cytokine receptor polypeptide encoded by the polynucleotide forms a heterodimeric or multimeric receptor complex. In another embodiment, the polynucleotide is as disclosed above, wherein the soluble cytokine receptor polypeptide encoded by the polynucleotide further comprises a soluble Class I or Class II cytokine receptor. In another embodiment, the polynucleotide is as disclosed above, wherein the soluble cytokine receptor polypeptide encoded by the polynucleotide forms a heterodimeric or multimeric receptor complex further comprising a soluble CRF2-4 receptor polypeptide (SEQ ID NO:33) or a soluble IL-10 receptor polypeptide (SEQ ID NO:34), or a soluble DIRS1 receptor polypeptide (SEQ ID NO:35). In another embodiment, the polynucleotide is as disclosed above, wherein the soluble cytokine receptor polypeptide further encodes an intracellular domain. In another embodiment, the polynucleotide is as disclosed above, wherein the soluble cytokine receptor polypeptide further comprises an affinity tag.

Within another aspect, the present invention provides an expression vector comprising the following operably linked elements: (a) a transcription promoter; a first DNA segment encoding a soluble cytokine receptor polypeptide having an amino acid sequence as shown in SEQ ID NO:3; and a transcription terminator; and (b) a second transcription promoter; a second DNA segment encoding a soluble Class I or Class II cytokine receptor polypeptide; and a transcription terminator; and wherein the first and second DNA segments are contained within a single expression vector or are contained within independent expression vectors.

In one embodiment, the expression vector disclosed above, further comprises a secretory signal sequence operably linked to the first and second DNA segments. In another embodiment, the expression vector is as disclosed above, wherein the second DNA segment encodes a soluble CRF2-4 receptor polypeptide (SEQ ID NO:33) or a soluble IL-10 receptor polypeptide (SEQ ID NO:34), or a soluble DIRS1 receptor polypeptide (SEQ ID NO:35).

Within another aspect, the present invention provides a cultured cell comprising an expression vector as disclosed above, wherein the cell expresses the polypeptides encoded by the DNA segments. In one embodiment, the cultured cell comprises an expression vector as disclosed above, wherein the first and second DNA segments are located on independent expression vectors and are co-transfected into the cell, and cell expresses the polypeptides encoded by the DNA segments. In another embodiment, the cultured cell comprises an expression vector as disclosed above, wherein the cell expresses a heterodimeric or multimeric soluble receptor polypeptide encoded by the DNA segments. In another embodiment, the cultured cell is as disclosed above, wherein the cell secretes a soluble cytokine receptor polypeptide heterodimer or multimeric complex. In another embodiment, the cultured cell is as disclosed above, wherein the cell secretes a soluble cytokine receptor polypeptide heterodimer or multimeric complex that binds IL-TIF or antagonizes IL-TIF activity.

Within another aspect, the present invention provides a DNA construct encoding a fusion protein comprising: a first DNA segment encoding a polypeptide having a sequence of amino acid residues as shown in SEQ ID NO:3; and at least one other DNA segment encoding a soluble Class I or Class II cytokine receptor polypeptide, wherein the first and other DNA segments are connected in-frame; and wherein the first and other DNA segments encode the fusion protein.

Within another aspect, the present invention provides a DNA construct encoding a fusion protein as disclosed above, wherein at least one other DNA segment encodes a soluble CRF2-4 receptor polypeptide (SEQ ID NO:33) or a soluble IL-10 receptor polypeptide (SEQ ID NO:34), or a soluble DIRS1 receptor polypeptide (SEQ ID NO:35).

Within another aspect, the present invention provides an expression vector comprising the following operably linked elements: a transcription promoter; a DNA construct encoding a fusion protein as disclosed above; and

a transcription terminator, wherein the promoter is operably linked to the DNA construct, and the DNA construct is operably linked to the transcription terminator.

Within another aspect, the present invention provides a cultured cell comprising an expression vector as disclosed above, wherein the cell expresses a polypeptide encoded by the DNA construct.

5 Within another aspect, the present invention provides a method of producing a fusion protein comprising: culturing a cell as disclosed above; and isolating the polypeptide produced by the cell.

 Within another aspect, the present invention provides an isolated soluble cytokine receptor polypeptide comprising a sequence of amino acid residues that is at least 90% identical to an amino acid sequence as shown in SEQ ID NO:3, and wherein
10 the soluble cytokine receptor polypeptide binds IL-TIF or antagonizes IL-TIF activity. In one embodiment, the isolated polypeptide is as disclosed above, wherein the soluble cytokine receptor polypeptide forms a homodimeric receptor complex.

 Within another aspect, the present invention provides an isolated polypeptide comprising a sequence of amino acid residues that is at least 90% identical
15 to an amino acid sequence as shown in SEQ ID NO:3, wherein the soluble cytokine receptor polypeptide forms a heterodimeric or multimeric receptor complex. In one embodiment, the isolated polypeptide is as disclosed above, wherein the soluble cytokine receptor polypeptide forms a heterodimeric or multimeric receptor complex further comprising a soluble Class I or Class II cytokine receptor.

20 In another embodiment, the isolated polypeptide is as disclosed above, wherein the soluble cytokine receptor polypeptide forms a heterodimeric or multimeric receptor complex further comprising a soluble CRF2-4 receptor polypeptide (SEQ ID NO:33) or a soluble IL-10 receptor polypeptide (SEQ ID NO:34), or a soluble DIRS1 receptor polypeptide (SEQ ID NO:35). In another embodiment, the isolated polypeptide
25 is as disclosed above, wherein the polypeptide forms a heterodimeric or multimeric receptor complex further comprising a soluble CRF2-4 receptor polypeptide (SEQ ID NO:33) or a soluble IL-10 receptor polypeptide (SEQ ID NO:34), or a soluble DIRS1 receptor polypeptide (SEQ ID NO:35).

 Within another aspect, the present invention provides an isolated soluble
30 cytokine receptor polypeptide comprising a sequence of amino acid residues as shown in SEQ ID NO:3, wherein the soluble cytokine receptor polypeptide forms a

heterodimeric or multimeric receptor complex. In another embodiment, the isolated polypeptide is as disclosed above, wherein the soluble cytokine receptor polypeptide forms a heterodimeric or multimeric receptor complex further comprising a soluble Class I or Class II cytokine receptor. In another embodiment, the isolated polypeptide is as disclosed above, wherein the soluble cytokine receptor polypeptide forms a heterodimeric or multimeric receptor complex comprising a soluble CRF2-4 receptor polypeptide (SEQ ID NO:33) or a soluble IL-10 receptor polypeptide (SEQ ID NO:34), or a soluble DIRS1 receptor polypeptide (SEQ ID NO:35). In another embodiment, the isolated polypeptide is as disclosed above, wherein the soluble cytokine receptor polypeptide further comprises an affinity tag, chemical moiety, toxin, or label. Within another aspect, the present invention provides an isolated heterodimeric or multimeric soluble receptor complex comprising soluble receptor subunits, wherein at least one of soluble receptor subunits comprises a soluble cytokine receptor polypeptide comprising a sequence of amino acid residues as shown in SEQ ID NO:3. In one embodiment, the isolated heterodimeric or multimeric soluble receptor complex disclosed above, further comprises a soluble Class I or Class II cytokine receptor polypeptide. In one embodiment, the isolated heterodimeric or multimeric soluble receptor complex disclosed above, further comprises a soluble CRF2-4 receptor polypeptide (SEQ ID NO:33) or a soluble IL-10 receptor polypeptide (SEQ ID NO:34), or a soluble DIRS1 receptor polypeptide (SEQ ID NO:35).

Within another aspect, the present invention provides a method of producing a soluble cytokine receptor polypeptide that forms a heterodimeric or multimeric complex comprising: culturing a cell as disclosed above; and isolating the soluble receptor polypeptides produced by the cell.

Within another aspect, the present invention provides a method of producing an antibody to soluble cytokine receptor polypeptide comprising: inoculating an animal with a soluble cytokine receptor polypeptide selected from the group consisting of: (a) a polypeptide comprising a homodimeric soluble cytokine receptor complex; (b) a polypeptide comprising a soluble cytokine receptor heterodimeric or multimeric receptor complex comprising a soluble Class I or Class II cytokine receptor polypeptide; (c) a polypeptide comprising a soluble cytokine

receptor heterodimeric or multimeric receptor complex comprising a soluble CRF2-4 receptor polypeptide (SEQ ID NO:33); (d) a polypeptide comprising a soluble cytokine receptor heterodimeric or multimeric receptor complex comprising a soluble IL-10 receptor polypeptide (SEQ ID NO:34); (e) a polypeptide comprising a soluble
5 cytokine receptor heterodimeric or multimeric receptor complex comprising a soluble DIRS1 receptor polypeptide (SEQ ID NO:34); and wherein the polypeptide elicits an immune response in the animal to produce the antibody; and isolating the antibody from the animal.

Within another aspect, the present invention provides an antibody
10 produced by the method as disclosed above, which specifically binds to a homodimeric, heterodimeric or multimeric receptor complex comprising a soluble cytokine receptor polypeptide. In one embodiment, the antibody disclosed above is a monoclonal antibody.

Within another aspect, the present invention provides an antibody which
15 specifically binds to a homodimeric, heterodimeric or multimeric receptor complex as disclosed above.

Within another aspect, the present invention provides a method for inhibiting IL-TIF-induced proliferation of hematopoietic cells and hematopoietic cell progenitors comprising culturing bone marrow or peripheral blood cells with a
20 composition comprising an amount of soluble cytokine receptor sufficient to reduce proliferation of the hematopoietic cells in the bone marrow or peripheral blood cells as compared to bone marrow or peripheral blood cells cultured in the absence of soluble cytokine receptor. In one embodiment, the method is as disclosed above, wherein the hematopoietic cells and hematopoietic progenitor cells are lymphoid cells. In one
25 embodiment, the method is as disclosed above, wherein the lymphoid cells are macrophages or T cells.

Within another aspect, the present invention provides a method of reducing IL-TIF-induced or IL-9 induced inflammation comprising administering to a mammal with inflammation an amount of a composition of soluble cytokine receptor
30 sufficient to reduce inflammation.

Within another aspect, the present invention provides a method of suppressing an immune response in a mammal exposed to an antigen or pathogen comprising: (1) determining a level of an antigen- or pathogen-specific antibody; (2) administering a composition comprising soluble cytokine receptor polypeptide in an acceptable pharmaceutical vehicle; (3) determining a post administration level of antigen- or pathogen-specific antibody; (4) comparing the level of antibody in step (1) to the level of antibody in step (3), wherein a lack of increase or a decrease in antibody level is indicative of suppressing an immune response.

These and other aspects of the invention will become evident upon reference to the following detailed description and the attached drawing.

The term "allelic variant" is used herein to denote any of two or more alternative forms of a gene occupying the same chromosomal locus. Allelic variation arises naturally through mutation, and may result in phenotypic polymorphism within populations. Gene mutations can be silent (no change in the encoded polypeptide) or may encode polypeptides having altered amino acid sequence. The term allelic variant is also used herein to denote a protein encoded by an allelic variant of a gene.

"Corresponding to", when used in reference to a nucleotide or amino acid sequence, indicates the position in a second sequence that aligns with the reference position when two sequences are optimally aligned.

The term "expression vector" is used to denote a DNA molecule, linear or circular, that comprises a segment encoding a polypeptide of interest operably linked to additional segments that provide for its transcription. Such additional segments include promoter and terminator sequences, and may also include one or more origins of replication, one or more selectable markers, an enhancer, a polyadenylation signal, etc. Expression vectors are generally derived from plasmid or viral DNA, or may contain elements of both.

The term "isolated", when applied to a polynucleotide, denotes that the polynucleotide has been removed from its natural genetic milieu and is thus free of

other extraneous or unwanted coding sequences, and is in a form suitable for use within genetically engineered protein production systems.

"Operably linked", when referring to DNA segments, indicates that the segments are arranged so that they function in concert for their intended purposes, e.g. transcription initiates in the promoter and proceeds through the coding segment to the terminator.

A "polynucleotide" is a single- or double-stranded polymer of deoxyribonucleotide or ribonucleotide bases read from the 5' to the 3' end. Polynucleotides include RNA and DNA, and may be isolated from natural sources, synthesized *in vitro*, or prepared from a combination of natural and synthetic molecules.

The term "promoter" is used herein for its art-recognized meaning to denote a portion of a gene containing DNA sequences that provide for the binding of RNA polymerase and initiation of transcription. Promoter sequences are commonly, but not always, found in the 5' non-coding regions of genes.

The term "receptor" is used herein to denote a cell-associated protein, or a polypeptide subunit of such a protein, that binds to a bioactive molecule (the "ligand") and mediates the effect of the ligand on the cell. Binding of ligand to receptor results in a conformational change in the receptor (and, in some cases, receptor multimerization, i.e., association of identical or different receptor subunits) that causes interactions between the effector domain(s) and other molecule(s) in the cell. These interactions in turn lead to alterations in the metabolism of the cell. Metabolic events that are linked to receptor-ligand interactions include gene transcription, phosphorylation, dephosphorylation, cell proliferation, increases in cyclic AMP production, mobilization of cellular calcium, mobilization of membrane lipids, cell adhesion, hydrolysis of inositol lipids and hydrolysis of phospholipids. The term "receptor polypeptide" is used to denote complete receptor polypeptide chains and portions thereof, including isolated functional domains (e.g., ligand-binding domains).

A "secretory signal sequence" is a DNA sequence that encodes a polypeptide (a "secretory peptide") that, as a component of a larger polypeptide, directs the larger polypeptide through a secretory pathway of a cell in which it is synthesized.

The larger polypeptide is commonly cleaved to remove the secretory peptide during transit through the secretory pathway.

A "soluble receptor" is a receptor polypeptide that is not bound to a cell membrane. Soluble receptors are most commonly ligand-binding receptor polypeptides that lack transmembrane and cytoplasmic domains. Soluble receptors can comprise additional amino acid residues, such as affinity tags that provide for purification of the polypeptide or provide sites for attachment of the polypeptide to a substrate, or immunoglobulin constant region sequences. Many cell-surface receptors have naturally occurring, soluble counterparts that are produced by proteolysis or translated from alternatively spliced mRNAs. Receptor polypeptides are said to be substantially free of transmembrane and intracellular polypeptide segments when they lack sufficient portions of these segments to provide membrane anchoring or signal transduction, respectively.

The term "splice variant" is used herein to denote alternative forms of RNA transcribed from a gene. Splice variation arises naturally through use of alternative splicing sites within a transcribed RNA molecule, or less commonly between separately transcribed RNA molecules, and may result in several mRNAs transcribed from the same gene. Splice variants may encode polypeptides having altered amino acid sequence. The term splice variant is also used herein to denote a protein encoded by a splice variant of an mRNA transcribed from a gene.

Molecular weights and lengths of polymers determined by imprecise analytical methods (e.g., gel electrophoresis) will be understood to be approximate values. When such a value is expressed as "about" X or "approximately" X, the stated value of X will be understood to be accurate to $\pm 10\%$.

Cytokine receptors subunits are characterized by a multi-domain structure comprising a ligand-binding domain and an effector domain that is typically involved in signal transduction. Multimeric cytokine receptors include homodimers (e.g., PDGF receptor $\alpha\alpha$ and $\beta\beta$ isoforms, erythropoietin receptor, MPL (thrombopoietin receptor), and G-CSF receptor), heterodimers whose subunits each have ligand-binding and effector domains (e.g., PDGF receptor $\alpha\beta$ isoform), and multimers having component subunits with disparate functions (e.g., IL-2, IL-3, IL-4,

IL-5, IL-6, IL-7, and GM-CSF receptors). Some receptor subunits are common to a plurality of receptors. For example, the AIC2B subunit, which cannot bind ligand on its own but includes an intracellular signal transduction domain, is a component of IL-3 and GM-CSF receptors. Many cytokine receptors can be placed into one of four related families on the basis of their structures and functions. Class I hematopoietic receptors, for example, are characterized by the presence of a domain containing conserved cysteine residues and the WSXWS motif. Additional domains, including protein kinase domains; fibronectin type III domains; and immunoglobulin domains, which are characterized by disulfide-bonded loops, are present in certain hematopoietic receptors. Cytokine receptor structure has been reviewed by Urdal, Ann. Reports Med. Chem. 26:221-228, 1991 and Cosman, Cytokine 5:95-106, 1993. It is generally believed that under selective pressure for organisms to acquire new biological functions, new receptor family members arose from duplication of existing receptor genes leading to the existence of multi-gene families. Family members thus contain vestiges of the ancestral gene, and these characteristic features can be exploited in the isolation and identification of additional family members.

Cell-surface cytokine receptors are further characterized by the presence of additional domains. These receptors are anchored in the cell membrane by a transmembrane domain characterized by a sequence of hydrophobic amino acid residues (typically about 21-25 residues), which is commonly flanked by positively charged residues (Lys or Arg). On the opposite end of the protein from the extracellular domain and separated from it by the transmembrane domain is an intracellular domain.

The Zcytor11 receptor is a class II cytokine receptor. These receptors usually bind to four-helix-bundle cytokines. Interleukin-10 and the interferons have receptors in this class (e.g., interferon-gamma alpha and beta chains and the interferon-alpha/beta receptor alpha and beta chains). Class II cytokine receptors are characterized by the presence of one or more cytokine receptor modules (CRM) in their extracellular domains. The CRMs of class II cytokine receptors are somewhat different than the better known CRMs of class I cytokine receptors. While the class II CRMs contain two type-III fibronectin-like domains, they differ in organization.

Zcytor11, like all known class II receptors except interferon-alpha/beta receptor alpha chain, has only a single class II CRM in its extracellular domain. Zcytor11 is a receptor for a helical cytokine of the interferon/IL-10 class. As was stated above, Zcytor11 is similar to the interferon α receptor α chain. Uze *et al. Cell* 60 255-264 (1996) Analysis of a human cDNA clone encoding Zcytor11 (SEQ ID NO:1) revealed an open reading frame encoding 574 amino acids (SEQ ID NO:2) comprising an extracellular ligand-binding domain of approximately 211 amino acid residues (residues 18-228 of SEQ ID NO:2; SEQ ID NO:3), a transmembrane domain of approximately 23 amino acid residues (residues 229-251 of SEQ ID NO:2), and an intracellular domain of approximately 313 amino acid residues (residues 252 to 574 of SEQ ID NO:2). Those skilled in the art will recognize that these domain boundaries are approximate and are based on alignments with known proteins and predictions of protein folding. Deletion of residues from the ends of the domains is possible.

Moreover, the zcytor11 receptor has been shown to bind a ligand called T-cell inducible Factor (TIF, or IL-TIF) (See, WIPO publication WO 00/24758; Dumontier et al., *J. Immunol.* 164:1814-1819, 2000; and Xie et al., *J. Biol. Chem.* manuscript in press M005304200). The human IL-TIF nucleotide sequence is represented in SEQ ID NO:7 and corresponding polypeptide sequence is shown in SEQ ID NO:8. Within preferred embodiments, the soluble receptor form of zcytor11, residues 18-228 of SEQ ID NO:2, (SEQ ID NO:3) is a homodimer, heterodimer, or multimer that antagonizes the effects of IL-TIF *in vivo*. Antibodies to such homodimer, heterodimer, or multimers also serve as antagonists of IL-TIF activity.

IL-TIF has been shown to be induced in the presence of IL-9, and is suspected to be involved in promoting Th1-type immune responses. IL-9 stimulates proliferation, activation, differentiation and/or induction of immune function in a variety of ways and is implicated in asthma, lung mastocytosis, and other diseases, as well as activate STAT pathways. Antagonists of IL-TIF or IL-9 function can have beneficial use against such human diseases. The present invention provides such novel antagonists of IL-TIF.

The present invention is based in part upon the discovery of a novel heterodimeric soluble receptor protein having the structure of a class II cytokine

receptor, and antibodies thereto. The heterodimeric soluble receptor includes at least one zcytor11 soluble receptor subunit, disclosed in the commonly owned US Patent No. 5,965,704. A second soluble receptor polypeptide included in the heterodimeric soluble receptor belongs to the receptor subfamily that includes Interleukin-10 (Liu Y et al, J Immunol. 152; 1821-1829, 1994 (IL-10R cDNA) (SEQ ID NO:34)), the interferons (e.g., interferon-gamma alpha and beta chains and the interferon-alpha/beta receptor alpha and beta chains), CRF2-4 (Genbank Accession No. Z17227; SEQ ID NO:33), and DIRS1 (WIPO Publication WO99/46379, Schering Corporation, 1999; SEQ ID NO:35). The zcytor11 receptor in conjunction with CRF2-4 and IL-10 Receptor was shown to signal JAK-STAT pathway in response to the natural ligand for the zcytor11 receptor, IL-TIF (Xie et al., supra.). According to the present invention, a heterodimeric soluble zcytor11 receptor, as exemplified by a preferred embodiment of a soluble zcytor11 receptor + soluble CRF2-4 receptor heterodimer (zcytor11/ CRF2-4), can act as a potent antagonist of the IL-TIF. Other embodiments include soluble heterodimer zcytor11/ IL-10R, zcytor11/IL-9R, and other class II receptor subunits, as well as multimeric receptors including but not limited to zcytor11/ CRF2-4/IL-10R, and zcytor11/ CRF2-4/IL-9R.

Analysis of the tissue distribution of the mRNA corresponding zcytor11 cDNA showed that mRNA level was highest in pancreas, followed by a much lower levels in thymus, colon, liver, skin, lung, kidney and small intestine. Thus, particular embodiments of the present invention are directed toward use of soluble zcytor11 heterodimers as antagonists in inflammatory and immune diseases or conditions such as pancreatitis, type I diabetes (IDDM), pancreatic cancer, pancreatitis, Graves Disease, inflammatory bowel disease (IBD), Crohn's Disease, colon and intestinal cancer, diverticulosis, autoimmune disease, sepsis, asthma, end-stage renal diseases, psoriasis, organ or bone marrow transplant; and where inhibition of inflammation, immune suppression, reduction of proliferation of hematopoietic, immune, inflammatory or lymphoid cells, macrophages, T-cells (including Th1 and Th2 cells), suppression of immune response to a pathogen or antigen, or inhibition of IL-TIF or IL-9 cytokine production is desired.

Moreover, antibodies recognizing zcytoR11, soluble zcytoR11/CRF2-4 heterodimers, and multimers described herein and/or soluble zcytoR11/CRF2-4 heterodimers, and multimers themselves are useful to:

1) Antagonize or block signaling via the IL-TIF receptors in the treatment of autoimmune diseases such as IDDM, multiple sclerosis (MS), systemic Lupus erythematosus (SLE), myasthenia gravis, rheumatoid arthritis, and IBD to prevent or inhibit signaling in immune cells (e.g. lymphocytes, monocytes, leukocytes) via zcytoR11 (Hughes C et al., *J. Immunol* 153: 3319-3325, 1994). Alternatively anti-soluble zcytoR11, anti-soluble zcytoR11/CRF2-4 heterodimer or multimer monoclonal antibody (Mab) can be used as an antagonist to deplete unwanted immune cells to treat autoimmune disease. Asthma, allergy and other atopic disease may be treated with an MAb against soluble zcytoR11 soluble zcytoR11/CRF2-4 heterodimers to inhibit the immune response or to deplete offending cells. Blocking or inhibiting signaling via zcytoR11, using the polypeptides and antibodies of the present invention, may also benefit diseases of the pancreas, kidney, pituitary and neuronal cells. IDDM, NIDDM, pancreatitis, and pancreatic carcinoma may benefit. ZcytoR11 may serve as a target for MAb therapy of cancer where an antagonizing MAb inhibits cancer growth and targets immune-mediated killing. (Holliger P, and Hoogenboom, H: *Nature Biotech.* 16: 1015-1016, 1998). Mabs to soluble zcytoR11, and soluble zcytoR11/CRF2-4 heterodimers and multimers may also be useful to treat nephropathies such as glomerulosclerosis, membranous neuropathy, amyloidosis (which also affects the kidney among other tissues), renal arteriosclerosis, glomerulonephritis of various origins, fibroproliferative diseases of the kidney, as well as kidney dysfunction associated with SLE, IDDM, type II diabetes (NIDDM), renal tumors and other diseases.

2) Agonize or initiate signaling via the IL-TIF receptors in the treatment of autoimmune diseases such as IDDM, MS, SLE, myasthenia gravis, rheumatoid arthritis, and IBD. Anti-soluble zcytoR11, anti-soluble zcytoR11/CRF2-4 heterodimers and multimer monoclonal antibodies may signal lymphocytes or other immune cells to differentiate, alter proliferation, or change production of cytokines or cell surface proteins that ameliorate autoimmunity. Specifically, modulation of a T-helper cell response to an alternate pattern of cytokine secretion may deviate an autoimmune

response to ameliorate disease (Smith JA et al., J. Immunol. 160:4841-4849, 1998). Similarly, agonistic Anti-soluble zcytoR11, anti-soluble zcytoR11/CRF2-4 heterodimers and multimer monoclonal antibodies may be used to signal, deplete and deviate immune cells involved in asthma, allergy and atopoid disease. Signaling via zcytoR11 may also benefit diseases of the pancreas, kidney, pituitary and neuronal cells. IDDM, NIDDM, pancreatitis, and pancreatic carcinoma may benefit. ZcytoR11 may serve as a target for MAb therapy of pancreatic cancer where a signaling MAb inhibits cancer growth and targets immune-mediated killing (Tutt, AL et al., J Immunol. 161: 3175-3185, 1998). Similarly renal cell carcinoma may be treated with monoclonal antibodies to zcytoR11-comprising soluble receptors of the present invention.

Soluble zcytoR11, soluble zcytoR11/CRF2-4 heterodimers and multimers described herein can be used to neutralize/block IL-TIF activity in the treatment of autoimmune disease, atopic disease, NIDDM, pancreatitis and kidney dysfunction as described above. A soluble form of zcytoR11 may be used to promote an antibody response mediated by Th cells and/or to promote the production of IL-4 or other cytokines by lymphocytes or other immune cells.

The soluble receptors of the present invention are useful as antagonists of the IL-TIF cytokine. Such antagonistic effects can be achieved by direct neutralization or binding of the IL-TIF. In addition to antagonistic uses, the soluble receptors of the present invention can bind IL-TIF and act as carrier proteins for the IL-TIF cytokine, in order to transport the Ligand to different tissues, organs, and cells within the body. As such, the soluble receptors of the present invention can be fused or coupled to molecules, polypeptides or chemical moieties that direct the soluble-receptor-Ligand complex to a specific site, such as a tissue, specific immune cell, or tumor. Thus, the soluble receptors of the present invention can be used to specifically direct the action of the IL-TIF. See, Cosman, D. Cytokine 5: 95-106, 1993; and Fernandez-Botran, R. Exp. Opin. Invest. Drugs 9:497-513, 2000.

Moreover, the soluble receptors of the present invention can be used to stabilize the IL-TIF, to increase the bio-availability, therapeutic longevity, and/or efficacy of the Ligand by stabilizing the Ligand from degradation or clearance, or by targeting the ligand to a site of action within the body. For example the naturally

occurring IL-6/soluble IL-6R complex stabilizes IL-6 and can signal through the gp130 receptor. See, Cosman, D. supra, and Fernandez-Botran, R. supra.

Within preferred embodiments of the invention the isolated polynucleotides will hybridize to similar sized regions of nucleotides SEQ ID NO:1
5 corresponding to SEQ ID NO:3 or a sequence complementary thereto, under stringent conditions. In general, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Typical stringent conditions
10 are those in which the salt concentration is at least about 0.02 M at pH 7 and the temperature is at least about 60°C. As previously noted, the isolated polynucleotides of the present invention include DNA and RNA. Methods for isolating DNA and RNA are well known in the art. It is generally preferred to isolate RNA from pancreas or prostate tissues although cDNA can also be prepared using RNA from other tissues or
15 isolated as genomic DNA. Total RNA can be prepared using guanidine HCl extraction followed by isolation by centrifugation in a CsCl gradient (Chirgwin *et al.*, *Biochemistry* 18:52-94, (1979)). Poly (A)⁺ RNA is prepared from total RNA using the method of Aviv and Leder *Proc. Natl. Acad. Sci. USA* 69:1408-1412, (1972). Complementary DNA (cDNA) is prepared from poly(A)⁺ RNA using known methods.
20 Polynucleotides encoding Zcytor11 polypeptides are then identified and isolated by, for example, hybridization or PCR.

Those skilled in the art will recognize that the sequences disclosed in SEQ ID NO:3 and the corresponding nucleotides of SEQ ID NO:1 and represent single
25 alleles of the human Zcytor11 receptor. Allelic variants of these sequences can be cloned by probing cDNA or genomic libraries from different individuals according to standard procedures.

The present invention further provides counterpart receptors and polynucleotides from other species ("species orthologs"). Of particular interest are Zcytor11 receptors from other mammalian species, including murine, porcine, ovine,
30 bovine, canine, feline, equine, and non-human primates. Species orthologs of the human Zcytor11 receptor can be cloned using information and compositions provided

by the present invention in combination with conventional cloning techniques. For example, a cDNA can be cloned using mRNA obtained from a tissue or cell type that expresses the receptor. Suitable sources of mRNA can be identified by probing Northern blots with probes designed from the sequences disclosed herein. A library is then prepared from mRNA of a positive tissue or cell line. A receptor-encoding cDNA can then be isolated by a variety of methods, such as by probing with a complete or partial cDNA of human and other primates or with one or more sets of degenerate probes based on the disclosed sequences. A cDNA can also be cloned using the polymerase chain reaction, or PCR (Mullis, U.S. Patent No. 4,683,202), using primers designed from the sequences disclosed herein. Within an additional method, the cDNA library can be used to transform or transfect host cells, and expression of the cDNA of interest can be detected with an antibody to the receptor. Similar techniques can also be applied to the isolation of genomic clones.

The present invention also provides isolated soluble monomeric, homodimeric, heterodimeric and multimeric receptor polypeptides that comprise at least one zcytor11 receptor subunit that is substantially homologous to the receptor polypeptide of SEQ ID NO:3. By "isolated" is meant a protein or polypeptide that is found in a condition other than its native environment, such as apart from blood and animal tissue. In a preferred form, the isolated polypeptide is substantially free of other polypeptides, particularly other polypeptides of animal origin. It is preferred to provide the polypeptides in a highly purified form, i.e. greater than 95% pure, more preferably greater than 99% pure. The term "substantially homologous" is used herein to denote polypeptides having 50%, preferably 60%, more preferably at least 80%, sequence identity to the sequences shown in SEQ ID NO:3. Such polypeptides will more preferably be at least 90% identical, and most preferably 95% or more identical to SEQ ID NO:3. Percent sequence identity is determined by conventional methods. See, for example, *Altschul et al., Bull. Math. Bio.* 48: 603-616, (1986) and Henikoff and Henikoff, *Proc. Natl. Acad. Sci. USA* 89:10915-10919, (1992). Briefly, two amino acid sequences are aligned to optimize the alignment scores using a gap opening penalty of 10, a gap extension penalty of 1, and the "blossom 62" scoring matrix of Henikoff and

Henikoff (*id.*) as shown in Table 2 (amino acids are indicated by the standard one-letter codes). The percent identity is then calculated as:

$$\begin{array}{l}
 \text{Total number of identical matches} \\
 5 \quad \frac{\text{-----}}{\text{[length of the longer sequence plus the}} \times 100 \\
 \quad \quad \quad \text{number of gaps introduced into the longer} \\
 \quad \quad \quad \text{sequence in order to align the two} \\
 \quad \quad \quad \text{sequences]}
 \end{array}$$

10

Table 2[illegible]

Sequence identity of polynucleotide molecules is determined by similar methods using a ratio as disclosed above.

Those skilled in the art appreciate that there are many established algorithms available to align two amino acid sequences. The "FASTA" similarity search algorithm of Pearson and Lipman is a suitable protein alignment method for
5 examining the level of identity shared by an amino acid sequence disclosed herein and the amino acid sequence of a putative variant ztrypl. The FASTA algorithm is described by Pearson and Lipman, Proc. Nat'l Acad. Sci. USA 85:2444 (1988), and by Pearson, Meth. Enzymol. 183:63 (1990).

10 Briefly, FASTA first characterizes sequence similarity by identifying regions shared by the query sequence (*e.g.*, SEQ ID NO:3) and a test sequence that have either the highest density of identities (if the ktup variable is 1) or pairs of identities (if ktup=2), without considering conservative amino acid substitutions, insertions, or deletions. The ten regions with the highest density of identities are then rescored by
15 comparing the similarity of all paired amino acids using an amino acid substitution matrix, and the ends of the regions are "trimmed" to include only those residues that contribute to the highest score. If there are several regions with scores greater than the "cutoff" value (calculated by a predetermined formula based upon the length of the sequence and the ktup value), then the trimmed initial regions are examined to
20 determine whether the regions can be joined to form an approximate alignment with gaps. Finally, the highest scoring regions of the two amino acid sequences are aligned using a modification of the Needleman-Wunsch-Sellers algorithm (Needleman and Wunsch, J. Mol. Biol. 48:444 (1970); Sellers, SIAM J. Appl. Math. 26:787 (1974)), which allows for amino acid insertions and deletions. Preferred parameters for FASTA
25 analysis are: ktup=1, gap opening penalty=10, gap extension penalty=1, and substitution matrix=BLOSUM62. These parameters can be introduced into a FASTA program by modifying the scoring matrix file ("SMATRIX"), as explained in Appendix 2 of Pearson, Meth. Enzymol., *supra.*

FASTA can also be used to determine the sequence identity of nucleic
30 acid molecules using a ratio as disclosed above. For nucleotide sequence comparisons,

the ktup value can range between one to six, preferably from three to six, most preferably three, with other FASTA program parameters set as default.

The BLOSUM62 table (Table 2) is an amino acid substitution matrix derived from about 2,000 local multiple alignments of protein sequence segments, representing highly conserved regions of more than 500 groups of related proteins (Henikoff and Henikoff, Proc. Nat'l Acad. Sci. USA 89:10915 (1992)). Accordingly, the BLOSUM62 substitution frequencies can be used to define conservative amino acid substitutions that may be introduced into the amino acid sequences of the present invention. Although it is possible to design amino acid substitutions based solely upon chemical properties (as discussed below), the language "conservative amino acid substitution" preferably refers to a substitution represented by a BLOSUM62 value of greater than -1. For example, an amino acid substitution is conservative if the substitution is characterized by a BLOSUM62 value of 0, 1, 2, or 3. According to this system, preferred conservative amino acid substitutions are characterized by a BLOSUM62 value of at least 1 (e.g., 1, 2 or 3), while more preferred conservative amino acid substitutions are characterized by a BLOSUM62 value of at least 2 (e.g., 2 or 3).

Substantially homologous proteins and polypeptides are characterized as having one or more amino acid substitutions, deletions or additions. These changes are preferably of a minor nature, that is conservative amino acid substitutions (see Table 3) and other substitutions that do not significantly affect the folding or activity of the protein or polypeptide; small deletions, typically of one to about 30 amino acids; and small amino- or carboxyl-terminal extensions, such as an amino-terminal methionine residue, a small linker peptide of up to about 20-25 residues, or a small extension that facilitates purification (an affinity tag), such as a poly-histidine tract, protein A (*Nilsson et al., EMBO J.* 4:1075, (1985); *Nilsson et al., Methods Enzymol.* 198:3, (1991)), glutathione S transferase (Smith and Johnson, *Gene* 67:31, 1988), or other antigenic epitope or binding domain. See, in general *Ford et al., Protein Expression and Purification* 2: 95-107, (1991). DNAs encoding affinity tags are available from commercial suppliers (e.g., Pharmacia Biotech, Piscataway, NJ).

Table 3

<u>Conservative amino acid substitutions</u>	
5	Basic: arginine
	lysine
	histidine
	Acidic: glutamic acid
	aspartic acid
10	Polar: glutamine
	asparagine
	Hydrophobic: leucine
	isoleucine
	valine
15	Aromatic: phenylalanine
	tryptophan
	tyrosine
	Small: glycine
	alanine
20	serine
	threonine
	methionine

Essential amino acids in the receptor polypeptides of the present invention can be identified according to procedures known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis (Cunningham and Wells, *Science* 244, 1081-1085, (1989); Bass *et al.*, *Proc. Natl. Acad. Sci. USA* 88:4498-4502, (1991)). In the latter technique, single alanine mutations are introduced at every residue in the molecule, and the resultant mutant molecules are tested for biological activity (e.g., ligand binding and signal transduction) to identify amino acid residues that are critical to the activity of the molecule. Sites of ligand-receptor interaction can also be determined by analysis of crystal structure as determined by such techniques as nuclear magnetic resonance, crystallography or photoaffinity labeling. See, for example, de Vos *et al.*, *Science* 255:306-312, (1992); Smith *et al.*, *J. Mol. Biol.* 224:899-904,

(1992); Wlodaver *et al.*, *FEBS Lett.* 309:59-64, (1992). The identities of essential amino acids can also be inferred from analysis of homologies with related receptors.

Multiple amino acid substitutions can be made and tested using known methods of mutagenesis and screening, such as those disclosed by Reidhaar-Olson and Sauer *Science* 241:53-57, (1988) or Bowie and Sauer *Proc. Natl. Acad. Sci. USA* 86:2152-2156, (1989). Briefly, these authors disclose methods for simultaneously randomizing two or more positions in a polypeptide, selecting for functional polypeptide, and then sequencing the mutagenized polypeptides to determine the spectrum of allowable substitutions at each position. Other methods that can be used include phage display *e.g.*, Lowman *et al.*, *Biochem.* 30:10832-10837, (1991); Ladner *et al.*, U.S. Patent No. 5,223,409; Huse, WIPO Publication WO 92/06204) and region-directed mutagenesis (Derbyshire *et al.*, *Gene* 46:145, (1986); Ner *et al.*, *DNA* 7:127, (1988)).

Mutagenesis methods as disclosed above can be combined with high-throughput screening methods to detect activity of cloned, mutagenized receptors in host cells. Preferred assays in this regard include cell proliferation assays and biosensor-based ligand-binding assays, which are described below. Mutagenized DNA molecules that encode active receptors or portions thereof (*e.g.*, ligand-binding fragments) can be recovered from the host cells and rapidly sequenced using modern equipment. These methods allow the rapid determination of the importance of individual amino acid residues in a polypeptide of interest, and can be applied to polypeptides of unknown structure.

Using the methods discussed above, one of ordinary skill in the art can prepare a variety of polypeptides that comprise a soluble receptor subunit that is substantially homologous to SEQ ID NO:3 or allelic variants thereof and retain the ligand-binding properties of the wild-type receptor. Such polypeptides may include additional amino acids from an extracellular ligand-binding domain of a Zcytor11 receptor as well as part or all of the transmembrane and intracellular domains. Such polypeptides may also include additional polypeptide segments as generally disclosed above.

The receptor polypeptides of the present invention, including soluble homodimeric, heterodimeric and multimeric receptors, full-length receptors, receptor fragments (e.g. ligand-binding fragments), and fusion polypeptides can be produced in genetically engineered host cells according to conventional techniques. Suitable host
5 cells are those cell types that can be transformed or transfected with exogenous DNA and grown in culture, and include bacteria, fungal cells, and cultured higher eukaryotic cells. Eukaryotic cells, particularly cultured cells of multicellular organisms, are preferred. Techniques for manipulating cloned DNA molecules and introducing exogenous DNA into a variety of host cells are disclosed by Sambrook *et al.*, *Molecular*
10 *Cloning: A Laboratory Manual*, 2nd ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, (1989), and Ausubel *et al.*, *ibid.*, which are incorporated herein by reference.

In general, a DNA sequence encoding a Zcytor11 soluble receptor polypeptide, or a DNA sequence encoding an additional subunit of a heterodimeric or
15 multimeric Zcytor11 soluble receptor, e.g., CRF2-4 or IL10R, polypeptide is operably linked to other genetic elements required for its expression, generally including a transcription promoter and terminator, within an expression vector. The vector will also commonly contain one or more selectable markers and one or more origins of replication, although those skilled in the art will recognize that within certain systems
20 selectable markers may be provided on separate vectors, and replication of the exogenous DNA may be provided by integration into the host cell genome. Selection of promoters, terminators, selectable markers, vectors and other elements is a matter of routine design within the level of ordinary skill in the art. Many such elements are described in the literature and are available through commercial suppliers. Multiple
25 components of a soluble receptor complex can be co-transfected on individual expression vectors or be contained in a single expression vector. Such techniques of expressing multiple components of protein complexes are well known in the art.

To direct a homodimeric, heterodimeric and multimeric Zcytor11 receptor polypeptide into the secretory pathway of a host cell, a secretory signal
30 sequence (also known as a leader sequence, prepro sequence or pre sequence) is provided in the expression vector. The secretory signal sequence may be that of the

receptor, or may be derived from another secreted protein (e.g., t-PA) or synthesized *de novo*. The secretory signal sequence is joined to the ZcytorII DNA sequence in the correct reading frame. Secretory signal sequences are commonly positioned 5' to the DNA sequence encoding the polypeptide of interest, although certain signal sequences
5 may be positioned elsewhere in the DNA sequence of interest (see, e.g., Welch *et al.*, U.S. Patent No. 5,037,743; Holland *et al.*, U.S. Patent No. 5,143,830).

Cultured mammalian cells are preferred hosts within the present invention. Methods for introducing exogenous DNA into mammalian host cells include calcium phosphate-mediated transfection (Wigler *et al.*, *Cell* 14:725, (1978); Corsaro
10 and Pearson, *Somatic Cell Genetics* 7:603, (1981); Graham and Van der Eb, *Virology* 52:456, (1973)), electroporation (Neumann *et al.*, *EMBO J.* 1:841-845, (1982)), DEAE-dextran mediated transfection (Ausubel *et al.*, eds., *Current Protocols in Molecular Biology*, (John Wiley and Sons, Inc., NY, 1987), and liposome-mediated transfection (Hawley-Nelson *et al.*, *Focus* 15:73, (1993); Ciccarone *et al.*, *Focus* 15:80, (1993)),
15 which are incorporated herein by reference. The production of recombinant polypeptides in cultured mammalian cells is disclosed, for example, by Levinson *et al.*, U.S. Patent No. 4,713,339; Hagen *et al.*, U.S. Patent No. 4,784,950; Palmiter *et al.*, U.S. Patent No. 4,579,821; and Ringold, U.S. Patent No. 4,656,134. Suitable cultured mammalian cells include the COS-1 (ATCC No. CRL 1650), COS-7 (ATCC No. CRL
20 1651), BHK (ATCC No. CRL 1632), BHK 570 (ATCC No. CRL 10314), 293 (ATCC No. CRL 1573; Graham *et al.*, *J. Gen. Virol.* 36:59-72, 1977) and Chinese hamster ovary (e.g. CHO-K1; ATCC No. CCL 61) cell lines. Additional suitable cell lines are known in the art and available from public depositories such as the American Type Culture Collection, Rockville, Maryland. In general, strong transcription promoters are
25 preferred, such as promoters from SV-40 or cytomegalovirus. See, e.g., U.S. Patent No. 4,956,288. Other suitable promoters include those from metallothionein genes (U.S. Patent Nos. 4,579,821 and 4,601,978) and the adenovirus major late promoter.

Drug selection is generally used to select for cultured mammalian cells into which foreign DNA has been inserted. Such cells are commonly referred to as
30 "transfectants". Cells that have been cultured in the presence of the selective agent and are able to pass the gene of interest to their progeny are referred to as "stable

transfectants." A preferred selectable marker is a gene encoding resistance to the antibiotic neomycin. Selection is carried out in the presence of a neomycin-type drug, such as G-418 or the like. Selection systems may also be used to increase the expression level of the gene of interest, a process referred to as "amplification."

- 5 Amplification is carried out by culturing transfectants in the presence of a low level of the selective agent and then increasing the amount of selective agent to select for cells that produce high levels of the products of the introduced genes. A preferred amplifiable selectable marker is dihydrofolate reductase, which confers resistance to methotrexate. Other drug resistance genes (e.g. hygromycin resistance, multi-drug
10 resistance, puromycin acetyltransferase) can also be used.

Other higher eukaryotic cells can also be used as hosts, including insect cells, plant cells and avian cells. Transformation of insect cells and production of foreign polypeptides therein is disclosed by Guarino et al., U.S. Patent No. 5,162,222; Bang et al., U.S. Patent No. 4,775,624; and WIPO publication WO 94/06463, which are
15 incorporated herein by reference. The use of *Agrobacterium rhizogenes* as a vector for expressing genes in plant cells has been reviewed by Sinkar et al., *J. Biosci. (Bangalore)* 11:47-58, (1987).

Fungal cells, including yeast cells, and particularly cells of the genus *Saccharomyces*, can also be used within the present invention, such as for producing
20 receptor fragments or polypeptide fusions. Methods for transforming yeast cells with exogenous DNA and producing recombinant polypeptides therefrom are disclosed by, for example, Kawasaki, U.S. Patent No. 4,599,311; Kawasaki et al., U.S. Patent No. 4,931,373; Brake, U.S. Patent No. 4,870,008; Welch et al., U.S. Patent No. 5,037,743; and Murray et al., U.S. Patent No. 4,845,075. Transformed cells are selected by
25 phenotype determined by the selectable marker, commonly drug resistance or the ability to grow in the absence of a particular nutrient (e.g., leucine). A preferred vector system for use in yeast is the POT1 vector system disclosed by Kawasaki et al. (U.S. Patent No. 4,931,373), which allows transformed cells to be selected by growth in glucose-containing media. Suitable promoters and terminators for use in yeast include those
30 from glycolytic enzyme genes (see, e.g., Kawasaki, U.S. Patent No. 4,599,311; Kingsman et al., U.S. Patent No. 4,615,974; and Bitter, U.S. Patent No. 4,977,092) and

alcohol dehydrogenase genes. See also U.S. Patents Nos. 4,990,446; 5,063,154; 5,139,936 and 4,661,454. Transformation systems for other yeasts, including *Hansenula polymorpha*, *Schizosaccharomyces pombe*, *Kluyveromyces lactis*, *Kluyveromyces fragilis*, *Ustilago maydis*, *Pichia pastoris*, *Pichia methanolica*, *Pichia guilliermondii* and *Candida maltosa* are known in the art. See, for example, Gleeson *et al.*, *J. Gen. Microbiol.* 132:3459-3465, (1986) and Cregg, U.S. Patent No. 4,882,279. *Aspergillus* cells may be utilized according to the methods of McKnight *et al.*, U.S. Patent No. 4,935,349. Methods for transforming *Acremonium chrysogenum* are disclosed by Sumino *et al.*, U.S. Patent No. 5,162,228. Methods for transforming *Neurospora* are disclosed by Lambowitz, U.S. Patent No. 4,486,533.

Transformed or transfected host cells are cultured according to conventional procedures in a culture medium containing nutrients and other components required for the growth of the chosen host cells. A variety of suitable media, including defined media and complex media, are known in the art and generally include a carbon source, a nitrogen source, essential amino acids, vitamins and minerals. Media may also contain such components as growth factors or serum, as required. The growth medium will generally select for cells containing the exogenously added DNA by, for example, drug selection or deficiency in an essential nutrient which is complemented by the selectable marker carried on the expression vector or co-transfected into the host cell.

Within one aspect of the present invention, a novel soluble receptor of the present invention is produced by a cultured cell, and the cell is used to screen for ligands for the receptor, including the natural ligand, IL-TIF, as well as agonists and antagonists of the natural ligand. To summarize this approach, a cDNA or gene encoding the receptor is combined with other genetic elements required for its expression (*e.g.*, a transcription promoter), and the resulting expression vector is inserted into a host cell. Cells that express the DNA and produce functional receptor are selected and used within a variety of screening systems. Each component of the homodimeric, heterodimeric and multimeric receptor complex can be expressed in the same cell.

Mammalian cells suitable for use in expressing Zcytor11 receptors and transducing a receptor-mediated signal include cells that express other receptor subunits which may form a functional complex with Zcytor11. These subunits may include those of the interferon receptor family or of other class II or class I cytokine receptors, e.g., CRF2-4, IL-10R, and IL-9R. It is also preferred to use a cell from the same species as the receptor to be expressed. Within a preferred embodiment, the cell is dependent upon an exogenously supplied hematopoietic growth factor for its proliferation. Preferred cell lines of this type are the human TF-1 cell line (ATCC number CRL-2003) and the AML-193 cell line (ATCC number CRL-9589), which are GM-CSF-dependent human leukemic cell lines and BaF3 (Palacios and Steinmetz, *Cell* 41: 727-734, (1985)) which is an IL-3 dependent murine pre-B cell line. Other cell lines include BHK, COS-1 and CHO cells.

Suitable host cells can be engineered to produce the necessary receptor subunits or other cellular component needed for the desired cellular response. This approach is advantageous because cell lines can be engineered to express receptor subunits from any species, thereby overcoming potential limitations arising from species specificity. Species orthologs of the human receptor cDNA can be cloned and used within cell lines from the same species, such as a mouse cDNA in the BaF3 cell line. Cell lines that are dependent upon one hematopoietic growth factor, such as GM-CSF or IL-3, can thus be engineered to become dependent upon IL-TIF.

Cells expressing functional receptor are used within screening assays. A variety of suitable assays are known in the art. These assays are based on the detection of a biological response in a target cell. One such assay is a cell proliferation assay. Cells are cultured in the presence or absence of a test compound, and cell proliferation is detected by, for example, measuring incorporation of tritiated thymidine or by colorimetric assay based on the metabolic breakdown of 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) (Mosman, *J. Immunol. Meth.* 65: 55-63, (1983)). An alternative assay format uses cells that are further engineered to express a reporter gene. The reporter gene is linked to a promoter element that is responsive to the receptor-linked pathway, and the assay detects activation of transcription of the reporter gene. A preferred promoter element in this regard is a serum response element.

or SRE. See, e.g., Shaw *et al.*, *Cell* 56:563-572, (1989). A preferred such reporter gene is a luciferase gene (de Wet *et al.*, *Mol. Cell. Biol.* 7:725, (1987)). Expression of the luciferase gene is detected by luminescence using methods known in the art (e.g., Baumgartner *et al.*, *J. Biol. Chem.* 269:29094-29101, (1994); Schenborn and Goiffin,
5 *Promega_Notes* 41:11, 1993). Luciferase activity assay kits are commercially available from, for example, Promega Corp., Madison, WI. Target cell lines of this type can be used to screen libraries of chemicals, cell-conditioned culture media, fungal broths, soil samples, water samples, and the like. For example, a bank of cell-conditioned media samples can be assayed on a target cell to identify cells that produce ligand. Positive
10 cells are then used to produce a cDNA library in a mammalian expression vector, which is divided into pools, transfected into host cells, and expressed. Media samples from the transfected cells are then assayed, with subsequent division of pools, re-transfection, subculturing, and re-assay of positive cells to isolate a cloned cDNA encoding the ligand.

15 A natural ligand for the Zcytor11 receptor can also be identified by mutagenizing a cell line expressing the full-length receptor and culturing it under conditions that select for autocrine growth. See WIPO publication WO 95/21930. Within a typical procedure, IL-3 dependent BaF3 cells expressing Zcytor11 and the necessary additional subunits are mutagenized, such as with 2-ethylmethanesulfonate
20 (EMS). The cells are then allowed to recover in the presence of IL-3, then transferred to a culture medium lacking IL-3 and IL-4. Surviving cells are screened for the production of a IL-TIF, such as by adding soluble receptor to the culture medium or by assaying conditioned media on wild-type BaF3 cells and BaF3 cells expressing the receptor. Using this method, cells and tissues expressing IL-TIF can be identified.

25 An additional screening approach provided by the present invention includes the use of hybrid receptor polypeptides. These hybrid polypeptides fall into two general classes. Within the first class, the intracellular domain of Zcytor11, comprising approximately residues 252 to 574 of SEQ ID NO:2, is joined to the ligand-binding domain of a second receptor. It is preferred that the second receptor be a
30 hematopoietic cytokine receptor, such as mpi receptor (Souyri *et al.*, *Cell* 63: 1137-1147, (1990)). The hybrid receptor will further comprise a transmembrane domain,

which may be derived from either receptor. A DNA construct encoding the hybrid receptor is then inserted into a host cell. Cells expressing the hybrid receptor are cultured in the presence of a ligand for the binding domain and assayed for a response. This system provides a means for analyzing signal transduction mediated by Zcytor11 while using readily available ligands. This system can also be used to determine if particular cell lines are capable of responding to signals transduced by Zcytor11 heterodimers and multimers of the present invention. A second class of hybrid receptor polypeptides comprise the extracellular (ligand-binding) domain of Zcytor11 (approximately residues 18 to 228 of SEQ ID NO:2; SEQ ID NO:3) with an intracellular domain of a second receptor, preferably a hematopoietic cytokine receptor, and a transmembrane domain. Hybrid zacytor11 heterodimers and multimers of the present invention receptors of this second class are expressed in cells known to be capable of responding to signals transduced by the second receptor. Together, these two classes of hybrid receptors enable the identification of a responsive cell type for the development of an assay for detecting a IL-TIF. Moreover, such cells can be used in the presence of IL-TIF to assay the soluble receptor antagonists of the present invention in a competition-type assay. In such assay, a decrease in the proliferation or signal transduction activity of IL-TIF in the presence of a soluble receptor of the present invention demonstrates antagonistic activity. Moreover IL-TIF-soluble receptor binding assays can be used to assess whether a soluble receptor antagonizes IL-TIF activity.

Cells found to express the ligand are then used to prepare a cDNA library from which the ligand-encoding cDNA can be isolated as disclosed above. The present invention thus provides, in addition to novel receptor polypeptides, methods for cloning polypeptide ligands for the receptors.

The tissue specificity of Zcytor11 expression suggests a role in the development of the pancreas, small intestine, colon and the thymus. In view of the tissue specificity observed for this receptor, agonists (including the natural ligand) and antagonists have enormous potential in both *in vitro* and *in vivo* applications. Compounds identified as receptor agonists are useful for stimulating proliferation and development of target cells *in vitro* and *in vivo*. For example, agonist compounds are

useful as components of defined cell culture media, and may be used alone or in combination with other cytokines and hormones to replace serum that is commonly used in cell culture. Agonists or antagonist may be useful in specifically regulating the growth and/or development of pancreatic, gastro-intestinal or thymic-derived cells in culture. These compounds are useful as research reagents for characterizing sites of ligand-receptor interaction. *In vivo*, receptor agonists or antagonists may find application in the treatment pancreatic, gastro-intestinal or thymic diseases.

Agonists or antagonists to Zcytor11 may include small families of peptides. These peptides may be identified employing affinity selection conditions that are known in the art, from a population of candidates present in a peptide library. Peptide libraries include combinatorial libraries chemically synthesized and presented on solid support (Lam *et al.*, *Nature* 354: 82-84 (1991)) or are in solution (Houghten *et al.*, *BioTechniques* 13: 412-421, (1992)), expressed then linked to plasmid DNA (Cull *et al.*, *Proc. Natl. Acad. Sci. USA* 89: 1865-1869 (1992)) or expressed and subsequently displayed on the surfaces of viruses or cells (Boder and Wittrup, *Nature Biotechnology* 15: 553-557(1997); Cwirla *et al. Science* 276: 1696-1699 (1997)).

Zcytor11 homodimeric, heterodimeric and multimeric may also be used within diagnostic systems for the detection of circulating levels of IL-TIF ligand. Within a related embodiment, antibodies or other agents that specifically bind to Zcytor11 soluble receptors of the present invention can be used to detect circulating receptor polypeptides. Elevated or depressed levels of ligand or receptor polypeptides may be indicative of pathological conditions, including cancer.

Zcytor11 homodimeric, heterodimeric and multimeric receptor polypeptides can be prepared by expressing a truncated DNA encoding the extracellular domain, for example, a polypeptide which contains SEQ ID NO:3 or the corresponding region of a non-human receptor. It is preferred that the extracellular domain polypeptides be prepared in a form substantially free of transmembrane and intracellular polypeptide segments. For example, the C-terminus of the receptor polypeptide may be at residue 228 of SEQ ID NO:2 or the corresponding region of an allelic variant or a non-human receptor. To direct the export of the receptor domain from the host cell, the receptor DNA is linked to a second DNA segment encoding a secretory peptide, such as

a t-PA secretory peptide. To facilitate purification of the secreted receptor domain, a C-terminal extension, such as a poly-histidine tag, substance P, FlagTM peptide {Hopp *et al.*, *Biotechnology* 6:1204-1210, (1988); available from Eastman Kodak Co., New Haven, CT) or another polypeptide or protein for which an antibody or other specific
5 binding agent is available, can be fused to the receptor polypeptide. Moreover, heterodimeric and multimeric non-zcytor11 subunit extracellular cytokine binding domains are also prepared as above.

In an alternative approach, a receptor extracellular domain of zcytor11 or other class I or II cytokine receptor component can be expressed as a fusion with
10 immunoglobulin heavy chain constant regions, typically an F_C fragment, which contains two constant region domains and a hinge region but lacks the variable region (See, Sledziewski, AZ et al., US Patent No. 6,018,026 and 5,750,375). The soluble zcytor11, soluble zcytoR11/CRF2-4 heterodimers and multimers of the present invention include such fusions. Such fusions are typically secreted as multimeric molecules wherein the
15 Fc portions are disulfide bonded to each other and two receptor polypeptides are arrayed in closed proximity to each other. Fusions of this type can be used to affinity purify the cognate ligand from solution, as an *in vitro* assay tool, to block signals *in vitro* by specifically titrating out ligand, and as antagonists *in vivo* by administering them parenterally to bind circulating ligand and clear it from the circulation. To purify
20 ligand, a Zcytor11-Ig chimera is added to a sample containing the ligand (e.g., cell-conditioned culture media or tissue extracts) under conditions that facilitate receptor-ligand binding (typically near-physiological temperature, pH, and ionic strength). The chimera-ligand complex is then separated by the mixture using protein A, which is immobilized on a solid support (e.g., insoluble resin beads). The ligand is then eluted
25 using conventional chemical techniques, such as with a salt or pH gradient. In the alternative, the chimera itself can be bound to a solid support, with binding and elution carried out as above. The chimeras may be used *in vivo* to regulate gastrointestinal, pancreatic or thymic functions. Chimeras with high binding affinity are administered parenterally (e.g., by intramuscular, subcutaneous or intravenous injection). Circulating
30 molecules bind ligand and are cleared from circulation by normal physiological

processes. For use in assays, the chimeras are bound to a support via the F_C region and used in an ELISA format.

The present invention further provides a variety of other polypeptide fusions and related multimeric proteins comprising one or more polypeptide fusions. For example, a soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 can be prepared as a fusion to a dimerizing protein as disclosed in U.S. Patents Nos. 5,155,027 and 5,567,584. Preferred dimerizing proteins in this regard include immunoglobulin constant region domains, e.g., IgGyl, and the human κ light chain. Immunoglobulin-soluble zcytor11 receptor or immunoglobulin-soluble zcytor11 heterodimeric polypeptide, such as immunoglobulin-soluble zcytor11/CRF2-4 fusions can be expressed in genetically engineered cells to produce a variety of multimeric zcytor11 receptor analogs. Auxiliary domains can be fused to soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 to target them to specific cells, tissues, or macromolecules (e.g., collagen, or cells expressing the IL-TIF). A zcytor11 polypeptide can be fused to two or more moieties, such as an affinity tag for purification and a targeting domain. Polypeptide fusions can also comprise one or more cleavage sites, particularly between domains. See, Tuan et al., Connective Tissue Research 34:1-9, 1996.

A preferred assay system employing a ligand-binding receptor fragment uses a commercially available biosensor instrument (BIAcore™, Pharmacia Biosensor, Piscataway, NJ), wherein the receptor fragment is immobilized onto the surface of a receptor chip. Use of this instrument is disclosed by Karlsson, *J. Immunol. Methods* 145:229-240, (1991) and Cunningham and Wells, *J. Mol. Biol.* 234:554-563, (1993). A receptor fragment is covalently attached, using amine or sulphydryl chemistry, to dextran fibers that are attached to gold film within the flow cell. A test sample is passed through the cell. If ligand is present in the sample, it will bind to the immobilized receptor polypeptide, causing a change in the refractive index of the medium, which is detected as a change in surface plasmon resonance of the gold film. This system allows the determination of on- and off-rates, from which binding affinity can be calculated, and assessment of stoichiometry of binding.

Ligand-binding receptor polypeptides can also be used within other assay systems known in the art. Such systems include Scatchard analysis for determination of binding affinity. See, Scatchard, *Ann. NY Acad. Sci.* 51: 660-672, (1949) and calorimetric assays (Cunningham *et al.*, *Science* 253:545-548, (1991);
5 Cunningham *et al.*, *Science* 254:821-825, (1991)).

A receptor ligand-binding polypeptide can also be used for purification of IL-TIF ligand. The receptor polypeptide is immobilized on a solid support, such as beads of agarose, cross-linked agarose, glass, cellulosic resins, silica-based resins, polystyrene, cross-linked polyacrylamide, or like materials that are stable under the
10 conditions of use. Methods for linking polypeptides to solid supports are known in the art, and include amine chemistry, cyanogen bromide activation, N-hydroxysuccinimide activation, epoxide activation, sulfhydryl activation, and hydrazide activation. The resulting media will generally be configured in the form of a column, and fluids containing ligand are passed through the column one or more times to allow ligand to
15 bind to the receptor polypeptide. The ligand is then eluted using changes in salt concentration or pH to disrupt ligand-receptor binding.

Moreover, soluble zcytor11 receptor or soluble zcytor11 heterodimeric receptor polypeptides, such as soluble zcytor11/CRF2-4, can be used as a "ligand sink," i.e., antagonist, to bind ligand *in vivo* or *in vitro* in therapeutic or other applications
20 where the presence of the ligand is not desired. For example, in cancers that are expressing large amounts of bioactive IL-TIF, soluble zcytor11 receptor or soluble zcytor11 heterodimeric and multimeric receptor polypeptides, such as soluble zcytor11/CRF2-4 can be used as a direct antagonist of the ligand *in vivo*, and may aid in reducing progression and symptoms associated with the disease, and can be used in
25 conjunction with other therapies (e.g., chemotherapy) to enhance the effect of the therapy in reducing progression and symptoms, and preventing relapse. Moreover, soluble zcytor11 receptor or soluble zcytor11 heterodimeric receptor polypeptides, such as soluble zcytor11/CRF2-4 can be used to slow the progression of cancers that over-express zcytor11 receptors, by binding ligand *in vivo* that would otherwise enhance
30 proliferation of those cancers.

Moreover, soluble zcytor11 receptor or soluble zcytor11 heterodimeric receptor polypeptides, such as soluble zcytor11/CRF2-4 can be used *in vivo* or in diagnostic applications to detect IL-TIF-expressing cancers *in vivo* or in tissue samples. For example, the soluble zcytor11 receptor or soluble zcytor11 heterodimeric receptor
5 polypeptides, such as soluble zcytor11/CRF2-4 can be conjugated to a radio-label or fluorescent label as described herein, and used to detect the presence of the IL-TIF in a tissue sample using an *in vitro* ligand-receptor type binding assay, or fluorescent imaging assay. Moreover, a radiolabeled soluble zcytor11 receptor or soluble zcytor11 heterodimeric receptor polypeptides, such as soluble zcytor11/CRF2-4 could be
10 administered *in vivo* to detect Ligand-expressing solid tumors through a radio-imaging method known in the art.

Soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 polypeptides can also be used to prepare antibodies that bind to epitopes, peptides, or polypeptides contained within the antigen. The
15 zcytor11 polypeptide or a fragment thereof serves as an antigen (immunogen) to inoculate an animal and elicit an immune response. One of skill in the art would recognize that antigens or immunogenic epitopes can consist of stretches of amino acids within a longer polypeptide, from about 10 amino acids and up to about the entire length of the polypeptide or longer depending on the polypeptide. Suitable antigens
20 include the zcytor11 polypeptide encoded by SEQ ID NO:3 or a contiguous 9 to 211 AA amino acid fragment thereof. Preferred peptides to use as antigens are the cytokine binding domain, disclosed herein, and zcytor11 hydrophilic peptides such as those predicted by one of skill in the art from a hydrophobicity plot, determined for example, from a Hopp/Woods hydrophilicity profile based on a sliding six-residue window, with
25 buried G, S, and T residues and exposed H, Y, and W residues ignored, or from a Jameson -Wolf plot of SEQ ID NO:3 using a DNA*STAR program. In addition, conserved motifs, and variable regions between conserved motifs of zcytor11 soluble receptor are suitable antigens. Suitable antigens also include the zcytor11 polypeptides disclosed above in combination with another class I or II cytokine extracellular domain,
30 such as those that form soluble zcytor11 heterodimeric polypeptides, such as soluble zcytor11/CRF2-4. Moreover, corresponding regions of the mouse soluble zcytor11

receptor polypeptide (SEQ ID NO:3) can be used to generate antibodies against the soluble mouse zcytor11 receptor. In addition Antibodies generated from this immune response can be isolated and purified as described herein. Methods for preparing and isolating polyclonal and monoclonal antibodies are well known in the art. See, for example, Current Protocols in Immunology, Cooligan, et al. (eds.), National Institutes of Health, John Wiley and Sons, Inc., 1995; Sambrook et al., Molecular Cloning: A Laboratory Manual, Second Edition, Cold Spring Harbor, NY, 1989; and Hurrell, J. G. R., Ed., Monoclonal Hybridoma Antibodies: Techniques and Applications, CRC Press, Inc., Boca Raton, FL, 1982.

As would be evident to one of ordinary skill in the art, polyclonal antibodies can be generated from inoculating a variety of warm-blooded animals such as horses, cows, goats, sheep, dogs, chickens, rabbits, mice, and rats with a soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 polypeptide or a fragment thereof. The immunogenicity of a zcytor11 polypeptide may be increased through the use of an adjuvant, such as alum (aluminum hydroxide) or Freund's complete or incomplete adjuvant. Polypeptides useful for immunization also include fusion polypeptides, such as fusions of zcytor11 or a portion thereof with an immunoglobulin polypeptide or with maltose binding protein. The polypeptide immunogen may be a full-length molecule or a portion thereof. If the polypeptide portion is "hapten-like", such portion may be advantageously joined or linked to a macromolecular carrier (such as keyhole limpet hemocyanin (KLH), bovine serum albumin (BSA) or tetanus toxoid) for immunization.

As used herein, the term "antibodies" includes polyclonal antibodies, affinity-purified polyclonal antibodies, monoclonal antibodies, and antigen-binding fragments, such as F(ab')₂ and Fab proteolytic fragments. Genetically engineered intact antibodies or fragments, such as chimeric antibodies, Fv fragments, single chain antibodies and the like, as well as synthetic antigen-binding peptides and polypeptides, are also included. Non-human antibodies may be humanized by grafting non-human CDRs onto human framework and constant regions, or by incorporating the entire non-human variable domains (optionally "cloaking" them with a human-like surface by replacement of exposed residues, wherein the result is a "veneered" antibody). In some

instances, humanized antibodies may retain non-human residues within the human variable region framework domains to enhance proper binding characteristics. Through humanizing antibodies, biological half-life may be increased, and the potential for adverse immune reactions upon administration to humans is reduced.

5 Antibodies are considered to be specifically binding if: 1) they exhibit a threshold level of binding activity, and 2) they do not significantly cross-react with related polypeptide molecules. A threshold level of binding is determined if anti-soluble zcytor11 receptor or anti-soluble zcytor11 heterodimeric polypeptide, such as anti-soluble zcytor11/CRF2-4 antibodies herein bind to a soluble zcytor11 receptor or
10 soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 polypeptide, peptide or epitope with an affinity at least 10-fold greater than the binding affinity to control (non-soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4) polypeptide. It is preferred that the antibodies exhibit a binding affinity (K_d) of 10^6 M^{-1} or greater, preferably 10^7 M^{-1} or
15 greater, more preferably 10^8 M^{-1} or greater, and most preferably 10^9 M^{-1} or greater. The binding affinity of an antibody can be readily determined by one of ordinary skill in the art, for example, by Scatchard analysis (Scatchard, G., Ann. NY Acad. Sci. 51: 660-672, 1949).

 Whether anti-soluble zcytor11 receptor or anti-soluble zcytor11
20 heterodimeric polypeptide, such as anti-soluble zcytor11/CRF2-4 antibodies do not significantly cross-react with related polypeptide molecules is shown, for example, by the antibody detecting soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 polypeptide but not known related polypeptides using a standard Western blot analysis (Ausubel et al., ibid.). Examples of
25 known related polypeptides are those disclosed in the prior art, such as known orthologs, and paralogs, and similar known members of a protein family. Screening can also be done using non-human soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4, and soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-
30 4 mutant polypeptides. Moreover, antibodies can be "screened against" known related polypeptides, to isolate a population that specifically binds to the soluble zcytor11

receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 polypeptides. For example, antibodies raised to soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 are adsorbed to related polypeptides adhered to insoluble matrix; antibodies specific to soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 will flow through the matrix under the proper buffer conditions. Screening allows isolation of polyclonal and monoclonal antibodies non-crossreactive to known closely related polypeptides (Antibodies: A Laboratory Manual, Harlow and Lane (eds.), Cold Spring Harbor Laboratory Press, 1988; Current Protocols in Immunology, Cooligan, et al. (eds.), National Institutes of Health, John Wiley and Sons, Inc., 1995). Screening and isolation of specific antibodies is well known in the art. See, Fundamental Immunology, Paul (eds.), Raven Press, 1993; Getzoff et al., *Adv. in Immunol.* 43: 1-98, 1988; Monoclonal Antibodies: Principles and Practice, Goding, J.W. (eds.), Academic Press Ltd., 1996; Benjamin et al., Ann. Rev. Immunol. 2: 67-101, 1984. Specifically binding anti-soluble zcytor11 receptor or anti-soluble zcytor11 heterodimeric polypeptide, such as anti-soluble zcytor11/CRF2-4 antibodies can be detected by a number of methods in the art, and disclosed below.

A variety of assays known to those skilled in the art can be utilized to detect antibodies that bind to soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 proteins or polypeptides. Exemplary assays are described in detail in Antibodies: A Laboratory Manual, Harlow and Lane (Eds.), Cold Spring Harbor Laboratory Press, 1988. Representative examples of such assays include: concurrent immunoelectrophoresis, radioimmunoassay, radioimmuno-precipitation, enzyme-linked immunosorbent assay (ELISA), dot blot or Western blot assay, inhibition or competition assay, and sandwich assay. In addition, antibodies can be screened for binding to wild-type versus mutant soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 protein or polypeptide.

Alternative techniques for generating or selecting antibodies useful herein include *in vitro* exposure of lymphocytes to soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 protein or

peptide, and selection of antibody display libraries in phage or similar vectors (for instance, through use of immobilized or labeled soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 protein or peptide). Genes encoding polypeptides having potential binding domains for soluble

5 zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 polypeptide, can be obtained by screening random peptide libraries displayed on phage (phage display) or on bacteria, such as *E. coli*. Nucleotide sequences encoding the polypeptides can be obtained in a number of ways, such as through random mutagenesis and random polynucleotide synthesis. These random

10 peptide display libraries can be used to screen for peptides which interact with a known target which can be a protein or polypeptide, such as a ligand or receptor, a biological or synthetic macromolecule, or organic or inorganic substances. Techniques for creating and screening such random peptide display libraries are known in the art (Ladner et al., US Patent NO. 5,223,409; Ladner et al., US Patent NO. 4,946,778;

15 Ladner et al., US Patent NO. 5,403,484 and Ladner et al., US Patent NO. 5,571,698) and random peptide display libraries and kits for screening such libraries are available commercially, for instance from Clontech (Palo Alto, CA), Invitrogen Inc. (San Diego, CA), New England Biolabs, Inc. (Beverly, MA) and Pharmacia LKB Biotechnology Inc. (Piscataway, NJ). Random peptide display libraries can be screened using the

20 soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 sequences disclosed herein to identify proteins which bind to soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4. These "binding polypeptides" which interact with soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble

25 zcytor11/CRF2-4 polypeptides can be used for tagging cells; for isolating homolog polypeptides by affinity purification; they can be directly or indirectly conjugated to drugs, toxins, radionuclides and the like. These binding polypeptides can also be used in analytical methods such as for screening expression libraries and neutralizing activity, e.g., for blocking interaction between IL-TIF ligand and receptor, or viral

30 binding to a receptor. The binding polypeptides can also be used for diagnostic assays for determining circulating levels of soluble zcytor11 receptor or soluble zcytor11

heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 polypeptides; for detecting or quantitating soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 polypeptides as marker of underlying pathology or disease. These binding polypeptides can also act as zcytor11 receptor or zcytor11
5 heterodimeric polypeptide, such as zcytor11/CRF2-4 "antagonists" to block zcytor11 receptor or zcytor11 heterodimeric polypeptide, such as zcytor11/CRF2-4 binding and signal transduction *in vitro* and *in vivo*. Again, these anti-soluble zcytor11 receptor or anti-soluble zcytor11 heterodimeric polypeptide, such as anti-soluble zcytor11/CRF2-4 binding polypeptides would be useful for inhibiting IL-TIF activity, as well as receptor
10 activity or protein-binding. Antibodies raised to the heterodimer or multimeric combinations of the present invention are preferred embodiments, as they may act more specifically against the IL-TIF, or more potently than antibodies raised to only one subunit. Moreover, the antagonistic and binding activity of the antibodies of the present invention can be assayed in the IL-TIF proliferation and other biological assays
15 described herein.

Antibodies to soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 may be used for tagging cells that express zcytor11 receptor or zcytor11 heterodimeric polypeptides, such as zcytor11/CRF2-4; for isolating soluble zcytor11 receptor or soluble zcytor11
20 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 polypeptide by affinity purification; for diagnostic assays for determining circulating levels of soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 polypeptides; for detecting or quantitating soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 as marker of
25 underlying pathology or disease; in analytical methods employing FACS; for screening expression libraries; for generating anti-idiotypic antibodies; and as neutralizing antibodies or as antagonists to block zcytor11 receptor or zcytor11 heterodimeric polypeptide, such as zcytor11/CRF2-4, or IL-TIF activity *in vitro* and *in vivo*. Suitable direct tags or labels include radionuclides, enzymes, substrates, cofactors, inhibitors,
30 fluorescent markers, chemiluminescent markers, magnetic particles and the like; indirect tags or labels may feature use of biotin-avidin or other complement/anti-

complement pairs as intermediates. Antibodies herein may also be directly or indirectly conjugated to drugs, toxins, radionuclides and the like, and these conjugates used for *in vivo* diagnostic or therapeutic applications. Moreover, antibodies to soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 or fragments thereof may be used *in vitro* to detect denatured or non-denatured soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 or fragments thereof in assays, for example, Western Blots or other assays known in the art.

Antibodies to soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 are useful for tagging cells that express the corresponding receptors and assaying their expression levels, for affinity purification, within diagnostic assays for determining circulating levels of soluble receptor polypeptides, analytical methods employing fluorescence-activated cell sorting. Moreover, divalent antibodies, and anti-idiotypic antibodies may be used as agonists to mimic the effect of the IL-TIF.

Antibodies herein can also be directly or indirectly conjugated to drugs, toxins, radionuclides and the like, and these conjugates used for *in vivo* diagnostic or therapeutic applications. For instance, antibodies or binding polypeptides which recognize soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 polypeptides of the present invention can be used to identify or treat tissues or organs that express a corresponding anti-complementary molecule (i.e., a zcytor11 receptor, or zcytor11 heterodimeric receptor, such as zcytor11/CRF2-4). More specifically, anti-soluble zcytor11 receptor or anti-soluble zcytor11 heterodimeric polypeptide, such as anti-soluble zcytor11/CRF2-4 antibodies, or bioactive fragments or portions thereof, can be coupled to detectable or cytotoxic molecules and delivered to a mammal having cells, tissues or organs that express the zcytor11 receptor or a zcytor11 heterodimeric receptor, such as zcytor11/CRF2-4 receptor molecules.

Suitable detectable molecules may be directly or indirectly attached to polypeptides that bind soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 ("binding polypeptides," including

binding peptides disclosed above), antibodies, or bioactive fragments or portions thereof. Suitable detectable molecules include radionuclides, enzymes, substrates, cofactors, inhibitors, fluorescent markers, chemiluminescent markers, magnetic particles and the like. Suitable cytotoxic molecules may be directly or indirectly
5 attached to the polypeptide or antibody, and include bacterial or plant toxins (for instance, diphtheria toxin, *Pseudomonas* exotoxin, ricin, abrin and the like), as well as therapeutic radionuclides, such as iodine-131, rhenium-188 or yttrium-90 (either directly attached to the polypeptide or antibody, or indirectly attached through means of a chelating moiety, for instance). Binding polypeptides or antibodies may also be
10 conjugated to cytotoxic drugs, such as adriamycin. For indirect attachment of a detectable or cytotoxic molecule, the detectable or cytotoxic molecule can be conjugated with a member of a complementary/ anticomplementary pair, where the other member is bound to the binding polypeptide or antibody portion. For these purposes, biotin/streptavidin is an exemplary complementary/ anticomplementary pair.

15 In another embodiment, binding polypeptide-toxin fusion proteins or antibody-toxin fusion proteins can be used for targeted cell or tissue inhibition or ablation (for instance, to treat cancer cells or tissues). Alternatively, if the binding polypeptide has multiple functional domains (i.e., an activation domain or a ligand binding domain, plus a targeting domain), a fusion protein including only the targeting
20 domain may be suitable for directing a detectable molecule, a cytotoxic molecule or a complementary molecule to a cell or tissue type of interest. In instances where the fusion protein including only a single domain includes a complementary molecule, the anti-complementary molecule can be conjugated to a detectable or cytotoxic molecule. Such domain-complementary molecule fusion proteins thus represent a generic
25 targeting vehicle for cell/tissue-specific delivery of generic anti-complementary-detectable/ cytotoxic molecule conjugates.

In another embodiment, soluble zcytor11 receptor or soluble zcytor11 heterodimeric polypeptide, such as soluble zcytor11/CRF2-4 binding polypeptide-cytokine or antibody-cytokine fusion proteins can be used for enhancing *in vivo* killing
30 of target tissues (for example, pancreatic, blood, lymphoid, colon, and bone marrow cancers), if the binding polypeptide-cytokine or anti-soluble zcytor11 receptor or anti-

soluble zcytor11 heterodimeric polypeptide, such as anti-soluble zcytor11/CRF2-4 antibody targets the hyperproliferative cell (See, generally, Hornick et al., Blood 89:4437-47, 1997). The described fusion proteins enable targeting of a cytokine to a desired site of action, thereby providing an elevated local concentration of cytokine.

- 5 Suitable anti-zcytor11 homodimer and heterodimer antibodies target an undesirable cell or tissue (i.e., a tumor or a leukemia), and the fused cytokine mediates improved target cell lysis by effector cells. Suitable cytokines for this purpose include interleukin 2 and granulocyte-macrophage colony-stimulating factor (GM-CSF), for instance.

Alternatively, soluble zcytor11 receptor or soluble zcytor11 heterodimeric
10 polypeptide, such as soluble zcytor11/CRF2-4 binding polypeptide or antibody fusion proteins described herein can be used for enhancing *in vivo* killing of target tissues by directly stimulating a zcytor11 receptor-modulated apoptotic pathway, resulting in cell death of hyperproliferative cells expressing zcytor11 receptor or a zcytor11 heterodimeric receptor, such as soluble zcytor11/CRF2-4 receptor.

15

The invention is further illustrated by the following non-limiting examples.

Example 1

20 Construction of Mammalian Expression Vectors That Express zcytor11 Soluble

Receptors: zcytor11CEE, zcytor11CFLG, zcytor11CHIS and zcytor11-Fc4

A. Construction of zcytor11 Mammalian Expression Vector containing zcytor11CEE, zcytor11CFLG and zcytor11CHIS

An expression vector is prepared for the expression of the soluble,
25 extracellular domain of the zcytor11 polypeptide (SEQ IDNO:3), pC4zcytor11CEE, wherein the construct is designed to express a zcytor11 polypeptide comprised of the predicted initiating methionine and truncated adjacent to the predicted transmembrane domain, and with a C-terminal Glu-Glu tag (SEQ ID NO:4).

A zcytor11 DNA fragment comprising the zcytor11 extracellular
30 cytokine binding domain (SEQ ID NO:3) is created using PCR, and purified. The excised DNA is subcloned into a plasmid expression vector that has a signal peptide,

e.g., the native zcytor11 signal peptide, and attaches a Glu-Glu tag (SEQ ID NO:4) to the C-terminus of the zcytor11 polypeptide-encoding polynucleotide sequence. Such an expression vector mammalian expression vector contains an expression cassette having a mammalian promoter, multiple restriction sites for insertion of coding sequences, a stop codon and a mammalian terminator. The plasmid can also have an *E. coli* origin of replication, a mammalian selectable marker expression unit having an SV40 promoter, enhancer and origin of replication, a DHFR gene and the SV40 terminator.

Restriction digested zcytor11 insert and previously digested vector are ligated using standard molecular biological techniques, and electroporated into competent cells such as DH10B competent cells (GIBCO BRL, Gaithersburg, MD) according to manufacturer's direction and plated onto LB plates containing 50 mg/ml ampicillin, and incubated overnight. Colonies are screened by restriction analysis of DNA prepared from individual colonies. The insert sequence of positive clones is verified by sequence analysis. A large scale plasmid preparation is done using a QIAGEN® Maxi prep kit (Qiagen) according to manufacturer's instructions.

The same process is used to prepare the zcytor11 soluble homodimeric, heterodimeric or multimeric receptors (including non-zcytor11 soluble receptor subunits, such as, soluble CRF2-4 or IL-10R) with a C-terminal his tag, composed of 6 His residues in a row; and a C-terminal flag (SEQ ID NO:5) tag, zcytor11CFLAG. To construct these constructs, the aforementioned vector has either the HIS or the FLAG® tag in place of the glu-glu tag (SEQ ID NO:4).

B. Mammalian Expression Construction of Soluble zcytor11 receptor zcytor11-Fc4

An expression plasmid containing all or part of a polynucleotide encoding zcytor11 is constructed via homologous recombination. A fragment of zcytor11 cDNA was isolated using PCR that includes the polynucleotide sequence from extracellular domain of the zcytor11 receptor. Primers used in PCR for the production of the zcytor11 fragment are from 5' to 3' end: (1) about 40 bp of the vector flanking sequence (5' of the insert) and about 17 bp corresponding to the 5' end of the zcytor11 extracellular domain; and (2) about 40 bp of the 5' end of the Fc4 polynucleotide sequence (SEQ ID NO:6) and about 17 bp corresponding to the 3' end of the zcytor11

extracellular domain. The fragment of Fc4 for fusion with the zcytorII is generated by PCR in a similar fashion. The two primers used in the production of the Fc4 fragment include: (1) a 5' primer consisting of about 40 bp of sequence from the 3' end of zcytorII extracellular domain and about 17 bp of the 5' end of Fc4 (SEQ ID NO:6);
5 and (2) a 3' primer consisting of about 40 bp of vector sequence (3' of the insert) and about 17 bp of the 3' end of Fc4 (SEQ ID NO:6). PCR amplification of the each of the reactions described above is then performed using conditions standard in the art.

An exemplary expression vector is derived from the plasmid pCZR199 (deposited at the American Type Culture Collection, 10801 University Boulevard,
10 Manassas, VA 20110-2209, designated No. 98668), that is cut with SmaI (BRL). The expression vector was derived from the plasmid pCZR199, and is a mammalian expression vector containing an expression cassette having the CMV immediate early promoter, a consensus intron from the variable region of mouse immunoglobulin heavy chain locus, multiple restriction sites for insertion of coding sequences, a stop codon
15 and a human growth hormone terminator. The expression vector also has an *E. coli* origin of replication, a mammalian selectable marker expression unit having an SV40 promoter, enhancer and origin of replication, a DHFR gene and the SV40 terminator. The expression vector used was constructed from pCZR199 by the replacement of the metallothionein promoter with the CMV immediate early promoter.

20 Competent yeast cells (*S. cerevisiae*) are combined with approximately 1 µg each of the zcytorII and Fc4 inserts, and 100 ng of SmaI (BRL) digested expression vector and electroporated. The yeast/DNA mixtures are electropulsed at, for example, 0.75 kV (5 kV/cm), "infinite" ohms, 25 µF. To each cuvette is added 600 µl of 1.2 M sorbitol and the yeast was plated in aliquots onto URA-D plates and incubated at 30°C.

25 After about 48 hours, the Ura+ yeast transformants from a single plate are picked, DNA isolated, and transformed into electrocompetent *E. coli* cells (e.g., DH10B, GibcoBRL), and plated using standard procedures. Individual clones harboring the correct expression construct for zcytorII-Fc4 are identified by restriction digest to verify the presence of the zcytorII-Fc4 insert and to confirm that the various
30 DNA sequences have been joined correctly to one another. The insert of positive

clones is subjected to sequence analysis. Larger scale plasmid DNA is isolated using the Qiagen Maxi kit (Qiagen) according to manufacturer's instructions.

Similar methods are used to prepare non-zcytor11 subunits of heterodimeric and multimeric receptors, such as CRF2-4 and IL-10R tagged with Fc4.

5

Example 2

Transfection And Expression of Soluble Receptor Polypeptides

BHK 570 cells (ATCC No. CRL-10314), DG-44 CHO, or other mammalian cells are plated at about 1.2×10^6 cells/well (6-well plate) in 800 μ l of appropriate serum free (SF) media (e.g., DMEM, Gibco/BRL High Glucose) (Gibco BRL, Gaithersburg, MD). The cells are transfected with expression plasmids containing zcytor11CEE, zcytor11CFLG, zcytor11CHIS or zcytor11-Fc4 (Example 1), or non-zcytor11 subunits of heterodimeric and multimeric receptors, such as -CEE, -CFLG, -CHIS, or -Fc4 tagged CRF2-4 and IL-10R, using Lipofectin™ (Gibco BRL), in serum free (SF) media according to manufacturer's instruction. Single clones expressing the soluble receptors are isolated, screened and grown up in cell culture media, and purified using standard techniques.

20

Example 3

Expression of zcytor11 Soluble Receptor in *E. coli*

A. Construction of expression vector pCZR225 that expresses huzcytor11/MBP-6H fusion polypeptide

An expression plasmid containing a polynucleotide encoding a zcytor11 soluble receptor fused C-terminally to maltose binding protein (MBP) is constructed via homologous recombination. The fusion polypeptide contains an N-terminal approximately 388 amino acid MBP portion fused to the zcytor11 soluble receptor (SEQ ID NO:3). A fragment of zcytor11 cDNA (SEQ ID NO:1) is isolated using PCR as described herein. Two primers are used in the production of the zcytor11 fragment in a standard PCR reaction: (1) one containing about 40 bp of the vector flanking sequence and about 25 bp corresponding to the amino terminus of the zcytor11, and (2) another containing about 40 bp of the 3' end corresponding to the flanking vector

30

sequence and about 25 bp corresponding to the carboxyl terminus of the zcytor11. Two μ l of the 100 μ l PCR reaction is run on a 1.0% agarose gel with 1 x TBE buffer for analysis, and the expected approximately fragment is seen. The remaining PCR reaction is combined with the second PCR tube and precipitated with 400 μ l of absolute ethanol. The precipitated DNA used for recombining into the SmaI cut recipient vector pTAP98 to produce the construct encoding the MBP-zcytor11 fusion, as described below.

Plasmid pTAP98 is derived from the plasmids pRS316 and pMAL-c2. The plasmid pRS316 is a *Saccharomyces cerevisiae* shuttle vector (Hieter P. and Sikorski, R., Genetics 122:19-27, 1989). pMAL-C2 (NEB) is an *E. coli* expression plasmid. It carries the tac promoter driving *MalE* (gene encoding MBP) followed by a His tag, a thrombin cleavage site, a cloning site, and the *rrnB* terminator. The vector pTAP98 is constructed using yeast homologous recombination. 100ng of EcoRI cut pMAL-c2 is recombined with 1 μ g PvuI cut pRS316, 1 μ g linker, and 1 μ g ScaI/EcoRI cut pRS316 are combined in a PCR reaction. PCR products are concentrated via 100% ethanol precipitation.

Competent yeast cells (*S. cerevisiae*) are combined with about 10 μ l of a mixture containing approximately 1 μ g of the zcytor11 receptor PCR product above, and 100 ng of SmaI digested pTAP98 vector, and electroporated using standard methods and plated onto URA-D plates and incubated at 30°C.

After about 48 hours, the Ura⁺ yeast transformants from a single plate are picked, DNA isolated, and transformed into electrocompetent *E. coli* cells (e.g., MC1061, Casadaban et. al. J. Mol. Biol. 138, 179-207), and plated on MM/CA +AMP 100 mg/L plates (Pryor and Leiting, Protein Expression and Purification 10:309-319, 1997).using standard procedures. Cells are grown in MM/CA with 100 μ g/ml Ampicillin for two hours, shaking, at 37°C. 1ml of the culture is induced with 1mM IPTG. 2-4 hours later the 250 μ l of each culture is mixed with 250 μ l acid washed glass beads and 250 μ l Thorner buffer with 5% β ME and dye (8M urea, 100 mM Tris pH7.0, 10% glycerol, 2mM EDTA, 5% SDS). Samples are vortexed for one minute and heated to 65°C for 10 minutes. 20 μ l are loaded per lane on a 4%-12% PAGE gel (NOVEX).

Gels are run in IXMES buffer. The positive clones are designated pCZR225 and subjected to sequence analysis.

One microliter of sequencing DNA is used to transform strain BL21. The cells are electropulsed at 2.0 kV, 25 μ F and 400 ohms. Following electroporation,
5 0.6 ml MM/CA with 100 mg/L Ampicillin. Cells are grown in MM/CA and induced with ITPG as described above., The positive clones are used to grow up for protein purification of the huzcytor11/MBP-6H fusion protein using standard techniques.

Example 4

10 Zcytor11 Soluble Receptor Polyclonal Antibodies

Polyclonal antibodies are prepared by immunizing female New Zealand white rabbits with the purified huzcytor11/MBP-6H polypeptide (Example 3), or the purified recombinant zcytor11CEE soluble receptor (Example 1). The rabbits are each given an initial intraperitoneal (IP) injection of 200 mg of purified protein in Complete
15 Freund's Adjuvant (Pierce, Rockford, IL) followed by booster IP injections of 100 mg purified protein in Incomplete Freund's Adjuvant every three weeks. Seven to ten days after the administration of the third booster injection, the animals are bled and the serum is collected. The rabbits are then boosted and bled every three weeks.

The zcytor11-specific polyclonal antibodies are affinity purified from the
20 rabbit serum using an CNBr-SEPHAROSE 4B protein column (Pharmacia LKB) that is prepared using about 10 mg of the purified huzcytor11/MBP-6H polypeptide per gram CNBr-SEPHAROSE, followed by 20X dialysis in PBS overnight. Zcytor11-specific antibodies are characterized by an ELISA titer check using 1 mg/ml of the appropriate protein antigen as an antibody target. The lower limit of detection (LLD) of the rabbit
25 anti-zcytor11 affinity purified antibodies is determined using standard methods.

Example 5

Zcytor11 Receptor Monoclonal Antibodies

Zcytor11 soluble receptor Monoclonal antibodies are prepared by
30 immunizing male BalbC mice (Harlan Sprague Dawley, Indianapolis, IN) with the purified recombinant soluble zcytor11 proteins described herein. The mice are each

given an initial intraperitoneal (IP) injection of 20 mg of purified protein in Complete Freund's Adjuvant (Pierce, Rockford, IL) followed by booster IP injections of 10 mg purified protein in Incomplete Freund's Adjuvant every two weeks. Seven to ten days after the administration of the third booster injection, the animals are bled and the
5 serum is collected, and antibody titer assessed.

Splenocytes are harvested from high-titer mice and fused to murine SP2/0 myeloma cells using PEG 1500 (Boehringer Mannheim, UK) in two separate fusion procedures using a 4:1 fusion ratio of splenocytes to myeloma cells (Antibodies: A Laboratory Manual, E. Harlow and D. Lane, Cold Spring Harbor Press). Following
10 10 days growth post-fusion, specific antibody-producing hybridomas are identified by ELISA using purified recombinant zcytor11 soluble receptor protein (Example 6C) as an antibody target and by FACS using Baf3 cells expressing the zcytor11 sequence (Example 8) as an antibody target. The resulting hybridomas positive by both methods are cloned three times by limiting dilution.

15

Example 6

Assessing Zcytor11 Receptor Heterodimerization Using ORIGIN Assay

Soluble zcytor11 receptor zcytor11CFLAG (Example 1), or gp130 (Hibi, M. et al., Cell 63:1149-1157, 1990) are biotinylated by reaction with a five-fold molar
20 excess of sulfo-NHS-LC-Biotin (Pierce, Inc., Rockford, IL) according to the manufacturer's protocol. Soluble zcytor11 receptor and another soluble receptor subunit, for example, soluble IL-10R (sIL-10R) or CRF2-4 receptor (CRF2-4) (R&D Systems, Minneapolis, MN), or soluble zcytor11 receptor (US Pat. No. 5,965,704) are labeled with a five fold molar excess of Ru-BPY-NHS (Igen, Inc., Gaithersburg, MD)
25 according to manufacturer's protocol. The biotinylated and Ru-BPY-NHS-labeled forms of the soluble zcytor11 receptor can be respectively designated Bio-zcytor11 receptor and Ru-zcytor11; the biotinylated and Ru-BPY-NHS-labeled forms of the other soluble receptor subunit can be similarly designated. Assays can be carried out using conditioned media from cells expressing a ligand, such as IL-TIF, that binds
30 zcytor11 heterodimeric receptors, or using purified IL-TIF.

For initial receptor binding characterization a panel of cytokines or conditioned medium are tested to determine whether they can mediate homodimerization of zcytor11 receptor and if they can mediate the heterodimerization of zcytor11 receptor with the soluble receptor subunits described above. To do this, 50 μ l of conditioned media or TBS-B containing purified cytokine, is combined with 50 μ l of TBS-B (20 mM Tris, 150 mM NaCl, 1 mg/ml BSA, pH 7.2) containing e.g., 400 ng/ml of Ru-zcytor11 receptor and Bio-zcytor11, or 400 ng/ml of Ru-zcytor11 receptor and e.g., Bio-gp130, or 400 ng/ml of e.g., Ru-CRF2-4 and Bio-zcytor11. Following incubation for one hour at room temperature, 30 μ g of streptavidin coated, 2.8 mm magnetic beads (Dyna, Inc., Oslo, Norway) are added and the reaction incubated an additional hour at room temperature. 200 μ l ORIGEN assay buffer (Igen, Inc., Gaithersburg, MD) is then added and the extent of receptor association measured using an M8 ORIGEN analyzer (Igen, Inc.).

Example 7

Construct for Generating a zcytor11 Receptor Heterodimer

A vector expressing a secreted human zcytor11 heterodimer was constructed. In this construct, the extracellular cytokine-binding domain of zcytor11 was fused to the heavy chain of IgG gamma 1 (IgGγ1) with a Glu-Glu tag (SEQ ID NO:4) at the C-terminus, while the extracellular portion of the heteromeric cytokine receptor subunit (e.g., an CRF2-4, IL-9, IL-10 IL-4 receptor component) was fused to the heavy chain of IgG gamma 1 (IgGγ1) with a His tag at the C-terminus.

A. Construction of IgG gamma 1 6-His and IgG gamma 1 Glu-Glu fusion vectors

The heavy chain of IgG γ l with a 6-His c-terminal tag (SEQ ID NO:13) was cloned into the pZP-9 mammalian expression vector (ATCC Deposit No. 98668) such that any desired cytokine receptor extracellular domain having a 5' EcoRI and 3' BamHI site can be cloned in, resulting in an N-terminal extracellular domain-C-terminal IgG γ l fusion. The IgG γ l fragment used in this construct was made by using PCR to isolate the IgG γ l using oligonucleotide primers ZC29,239 (SEQ ID NO:14) and ZC29,232 (SEQ ID NO:15). PCR products were purified using methods described

herein and digested with XhoI and BamHI (Boehringer-Mannheim) and subsequently gel purified. The extracellular portion of CRF2-4 (SEQ ID NO:18) was amplified using PCR, using oligonucleotide primers ZC39,319 (SEQ ID NO:16) and ZC39,325 (SEQ ID NO:17). PCR products were purified using methods described herein and digested with EcoRI and BamHI (Boehringer-Mannheim) and subsequently gel purified. The BamHI/XhoI IgGyl fragment and EcoRI/BamHI CRF2-4 fragment derived above were then ligated together into pZP-9 previously digested with EcoRI and XhoI to derive a construct that had the extracellular portion of CRF2-4 extracellular cytokine binding domain fused to IgGyl with a 6-HIS tag at the c-terminus. This construct was subsequently modified to introduce a thrombin cleavage site 3' of CRF2-4 and 5' of the 6-HIS tag. This was done by using the above construct as template in PCR with oligonucleotide primers ZC38,981 (SEQ ID NO:20) and ZC39,042 (SEQ ID NO:21). PCR products were purified using methods described herein and digested with SacII and XhoI. This SacII/XhoI fragment was ligated into the construction described above that had been previously digested with SacII and XhoI. The polynucleotide sequence of the CRF2-4 extracellular cytokine binding domain fused to IgGyl with a 6-HIS tag is shown in SEQ ID NO:22 and the corresponding polypeptide sequence is shown in SEQ ID NO:23.

The heavy chain of IgGyl with a Glu-Glu c-terminal tag (SEQ ID NO:4) was cloned into the Zem228R mammalian expression vector (ATCC deposit No. 69446) such that any desired cytokine receptor extracellular domain having a 5' EcoRI site and a 3' BamHI site can be cloned in resulting in a N-terminal cytokine extracellular domain-C-terminal IgGyl fusion. The IgGyl fragment used in this construct was made by using PCR to isolate the IgGyl sequence using oligonucleotide primers ZC29238 (SEQ ID NO:24) and ZC29231 (SEQ ID NO:25). PCR products were purified using methods described herein and digested with XhoI and EcoRI (Boehringer-Mannheim) and subsequently gel purified. The extracellular portion of hzcytor11 (SEQ ID NO:1) was amplified using PCR using oligonucleotides ZC39335 (SEQ ID NO:26) and ZC28981 (SEQ ID NO:27). PCR products were purified using methods described herein and digested with EcoRI and BamHI (Boehringer-Mannheim) and subsequently gel purified. The BamHI/XhoI IgGyl fragment and EcoRI/BamHI

hzcylor11 fragments derived above were then ligated together into Zem228R previously digested with EcoRI and XhoI to derive a construct that had the extracellular domain of hzcylor11 fused to IgG γ 1 with a Glu-Glu epitope tag at the C-terminus. This construct was subsequently modified to introduce a thrombin cleavage site 3' of CRF2-4 and 5' of the 6-HIS tag. This was done by using the above construct as template in PCR with oligonucleotide primers ZC38981 (SEQ ID NO:27) and ZC39043 (SEQ ID NO:28). PCR products were purified using methods described herein and digested with SacII and XhoI. This SacII/XhoI fragment was ligated into the construction described above that had been previously digested with SacII and XhoI. The polynucleotide sequence of the hzcylor11 extracellular cytokine binding domain fused to IgG γ 1 with a Glu Glu tag is shown in SEQ ID NO:29 and the corresponding polypeptide sequence is shown in SEQ ID NO:30.

B. Co-expression of the zcylor11 and heterodimeric cytokine receptor subunit extracellular domain

16 μ g of each of vectors above, were co-transfected into mammalian cells, e.g., BHK-570 cells (ATCC No. CRL-10314) using LipofectaminePlus™ reagent (Gibco/BRL), as per manufacturer's instructions. The transfected cells were selected for 10 days in DMEM + 5%FBS (Gibco/BRL) containing 1 μ M of methotrexate (MTX) (Sigma, St. Louis, MO) and 0.5 mg/ml G418 (Gibco/BRL) for 10 days. The resulting pool of transfectants was selected again in 10 μ M of MTX and 0.5 mg/ml G418 for 10 days.

The resulting pool of doubly selected cells was used to generate zcylor11/CRF2-4 soluble receptor protein. Three Factories (Nunc, Denmark) of this pool were used to generate 10 L of serum free conditioned medium. This conditioned media was passed over a nickel column followed by a Glu-Glu column to purify heterodimers away from homodimers.

Example 8Determination of Receptor Subunits That Heterodimerize or
Multimerize With zcytor11 Receptor

Using standard methods described herein, The BaF3/MPL-zcytor11
5 chimera cells are transfected with an additional heterodimeric cytokine receptor subunit
serve as a bioassay cell line to measure signal transduction response of heterodimeric
zcytor11 receptor complexes to the luciferase reporter in the presence of TPO. In the
presence of TPO, the BaF3/MPL-zcytor11 cells do not signal, suggesting that zcytor11
receptor must heterodimerize to signal. Transfection of the BaF3/MPL-zcytor11 cell
10 line with and additional MPL-class I cytokine receptor fusion that signals in the
presence of the TPO ligand, determines which heterodimeric cytokine receptor subunits
are required for zcytor11 receptor signaling. Use of MPL-receptor fusions for this
purpose alleviates the requirement for the presence of a natural ligand for the zcytor11
receptor.

15 MPL-class I cytokine receptor fusions are made as per Example 5 using
the extracellular domain and transmembrane domains of the MPL receptor and the
intracellular signaling domain of the desired class I cytokine receptor. The BaF3/MPL-
zcytor11 bioassay cell line co-transfected with an individual MPL-class I cytokine
receptor fusions as per Example 6 to form a BaF3/MPL-zcytor11/MPL-class I cytokine
20 receptor cell line. Receptor complexes include but are not limited to zcytor11 receptor
in combination with an MPL-cytokine receptor fusion comprising one or more of e.g., a
CRF2-4, IL-9, IL-10 IL-4 receptor component. Each independent receptor complex cell
line is then assayed in the presence of TPO and proliferation measured using routine
methods (e.g., Alamar Blue assay). The BaF3/MPL-zcytor11 bioassay cell line serves as
25 a control for the background luciferase activity, and is thus used as a baseline to
compare signaling by the various receptor complex combinations. In addition, a
BaF3/MPL-class I cytokine receptor cell line can be constructed to control for MPL-
class I cytokine receptor homodimerization effects for those class I cytokine receptors
known to signal upon homodimerization. The TPO in the presence of the correct
30 receptor complex, is expected to increase proliferation of the BaF3/MPL-

zcytor11/MPL-class I cytokine receptor cell line approximately 5 fold over background or greater in the presence of TPO.

Similar proliferation assays use full-length zcytor11 (SEQ ID NO:2) to screen for additional non-zcytor11 subunits that signal heterodimeric and multimeric complexes. Cells expressing full-length zcytor11 (SEQ ID NO:2) are transfected with a non-zcytor11 subunit and assayed for proliferation in the presence of IL-TIF ligand. Cells expressing the components of zcytor11 heterodimeric and multimeric receptors should proliferate in the presence of IL-TIF.

Example 9

Reconstitution of zcytor11 Receptor *in vitro*

To identify components involved in the zcytor11-signaling complex, receptor reconstitution studies are performed as follows. BHK 570 cells (ATCC No. CRL-10314) transfected, using standard methods described herein, with a luciferase reporter mammalian expression vector plasmid serve as a bioassay cell line to measure signal transduction response from a transfected zcytor11 receptor complex to the luciferase reporter in the presence of IL-TIF. BHK cells do not endogenously express the zcytor11 receptor. An exemplary luciferase reporter mammalian expression vector is the KZ134 plasmid which was constructed with complementary oligonucleotides that contain STAT transcription factor binding elements from 4 genes. A modified c-fos Sis inducible element (m67SIE, or hSIE) (Sadowski, H. et al., Science 261:1739-1744, 1993), the p21 SIE1 from the p21 WAF1 gene (Chin, Y. et al., Science 272:719-722, 1996), the mammary gland response element of the β -casein gene (Schmitt-Ney, M. et al., Mol. Cell. Biol. 11:3745-3755, 1991), and a STAT inducible element of the Fcg RI gene, (Seidel, H. et al., Proc. Natl. Acad. Sci. 92:3041-3045, 1995). These oligonucleotides contain Asp718-XhoI compatible ends and were ligated, using standard methods, into a recipient firefly luciferase reporter vector with a c-fos promoter (Poulsen, L.K. et al., J. Biol. Chem. 273:6229-6232, 1998) digested with the same enzymes and containing a neomycin selectable marker. The KZ134 plasmid is used to stably transfect BHK, or BaF3 cells, using standard transfection and selection methods, to make a BHK/KZ134 or BaF3/KZ134 cell line respectively.

The bioassay cell line is transfected with zcytor11 receptor alone, or co-transfected with zcytor11 receptor along with one of a variety of other known receptor subunits. Receptor complexes include but are not limited to zcytor11 receptor only, various combinations of zcytor11 receptor with one or more of the CRF2-4, IL-9, IL-10
 5 IL-4 receptor components, class II cytokine receptor subunits, or the IL-2 receptor components (IL-2R α , IL-2R β , IL-2R γ), zcytor11 receptor with one or more of the IL-4/IL-13 receptor family receptor components (IL-4R α , IL-13R α , IL-13R α'), as well as other Interleukin receptors (e.g., IL-15 R α , IL-7R α , IL-9R α , IL-21R (zcytor11)). Each independent receptor complex cell line is then assayed in the presence of cytokine-
 10 conditioned media or purified cytokines and luciferase activity measured using routine methods. The untransfected bioassay cell line serves as a control for the background luciferase activity, and is thus used as a baseline to compare signaling by the various receptor complex combinations. The conditioned medium or cytokine that binds the zcytor10 receptor in the presence of the correct receptor complex, is expected to give a
 15 luciferase readout of approximately 5 fold over background or greater.

As an alternative, a similar assay can be performed wherein the Baf3/zcytor11-mpl and Baf3/zcytor11 cell lines are co-transfected as described above and proliferation measured.

20

Example 10

Construct for Generating CEE-tagged IL-TIF

Oligonucleotides were designed to generate a PCR fragment containing the Kozak sequence and the coding region for IL-TIF, without its stop codon. These oligonucleotides were designed with a KpnI site at the 5' end and a BamHI site at the 3'
 25 end to facilitate cloning into pHZ200-CEE, our standard vector for mammalian expression of C-terminal Glu-Glu tagged (SEQ ID NO:4) proteins. The pHZ200 vector contains an MT-1 promoter.

PCR reactions were carried out using Turbo Pfu polymerase (Stratagene) to amplify a IL-TIF cDNA fragment. About 20 ng human IL-TIF polynucleotide
 30 template (SEQ ID NO:7), and oligonucleotides ZC28590 (SEQ ID NO:9) and ZC28580 (SEQ ID NO:10) were used in the PCR reaction. PCR reaction conditions were as

follows: 95°C for 5 minutes; 30 cycles of 95°C for 60 seconds, 55°C for 60 seconds, and 72°C for 60 seconds; and 72°C for 10 minutes; followed by a 4°C hold. PCR products were separated by agarose gel electrophoresis and purified using a QiaQuick™ (Qiagen) gel extraction kit. The isolated, approximately 600 bp, DNA fragment was
5 digested with KpnI and BamHI (Boehringer-Mannheim), gel purified as above and ligated into pHZ200-CEE that was previously digested with KpnI and BamHI.

About one microliter of the ligation reaction was electroporated into DH10B ElectroMax™ competent cells (GIBCO BRL, Gaithersburg, MD) according to manufacturer's direction and plated onto LB plates containing 100 µg/ml ampicillin,
10 and incubated overnight. Colonies were picked and screened by PCR using oligonucleotides ZC28590 (SEQ ID NO:9) and ZC28580 (SEQ ID NO:10), with PCR conditions as described above. Clones containing inserts were then sequenced to confirm error-free IL-TIF inserts. Maxipreps of the correct pHZ200-IL-TIF-CEE construct, as verified by sequence analysis, were performed.

15

Example 11

Transfection And Expression Of IL-TIF Polypeptides

BHK 570 cells (ATCC No. CRL-10314), were plated at about 1.2×10^6 cells/well (6-well plate) in 800 µl of serum free (SF) DMEM media (DMEM,
20 Gibco/BRL High Glucose) (Gibco BRL, Gaithersburg, MD). The cells were transfected with an expression plasmid containing IL-TIF-CEE described above (Example 10), using Lipofectin™ (Gibco BRL), in serum free (SF) DMEM according to manufacturer's instructions.

The cells were incubated at 37°C for approximately five hours, then
25 transferred to separate 150 mm MAXI plates in a final volume of 30 ml DMEM/5% fetal bovine serum (FBS) (Hyclone, Logan, UT). The plates were incubated at 37°C, 5% CO₂, overnight and the DNA: Lipofectin™ mixture was replaced with selection media (5% FBS/DMEM with 1 µM methotrexate (MTX)) the next day.

Approximately 10-12 days post-transfection, colonies were mechanically
30 picked to 12-well plates in one ml of 5%FCS/DMEM with 5 µM MTX, then grown to confluence. Positive expressing clonal colonies Conditioned media samples were then

tested for expression levels via SDS-PAGE and Western analysis. A high-expressing clone was picked and expanded for ample generation of conditioned media for purification of the IL-TIF-CEE expressed by the cells (Example 12).

5

Example 12

Purification of IL-TIF-CEE polypeptide from BHK 570 cells

Unless otherwise noted, all operations were carried out at 4°C. The following procedure was used for purifying IL-TIF polypeptide containing C-terminal GluGlu (EE) tags (SEQ ID NO:4). Conditioned media from BHK cells expressing IL-TIF-CEE (Example 11) was concentrated with an Amicon S10Y3 spiral cartridge on a ProFlux A30. A Protease inhibitor solution was added to the concentrated conditioned media to final concentrations of 2.5 mM ethylenediaminetetraacetic acid (EDTA, Sigma Chemical Co. St. Louis, MO), 0.003 mM leupeptin (Boehringer-Mannheim, Indianapolis, IN), 0.001 mM pepstatin (Boehringer-Mannheim) and 0.4 mM Pefabloc (Boehringer-Mannheim). Samples were removed for analysis and the bulk volume was frozen at -80°C until the purification was started. Total target protein concentrations of the concentrated conditioned media were determined via SDS-PAGE and Western blot analysis with the anti-EE HRP conjugated antibody.

About 100 ml column of anti-EE G-Sepharose (prepared as described below) was poured in a Waters AP-5, 5 cm x 10 cm glass column. The column was flow packed and equilibrated on a BioCad Sprint (PerSeptive BioSystems, Framingham, MA) with phosphate buffered saline (PBS) pH 7.4. The concentrated conditioned media was thawed, 0.2 micron sterile filtered, pH adjusted to 7.4, then loaded on the column overnight with about 1 ml/minute flow rate. The column was washed with 10 column volumes (CVs) of phosphate buffered saline (PBS, pH 7.4), then plug eluted with 200 ml of PBS (pH 6.0) containing 0.5 mg/ml EE peptide (Anaspec, San Jose, CA) at 5 ml/minute. The EE peptide used has the sequence EYMPME (SEQ ID NO:4). The column was washed for 10 CVs with PBS, then eluted with 5 CVs of 0.2M glycine, pH 3.0. The pH of the glycine-eluted column was adjusted to 7.0 with 2 CVs of 5X PBS, then equilibrated in PBS (pH 7.4). Five ml fractions were collected over the entire elution chromatography and absorbance at 280

and 215 nM were monitored; the pass through and wash pools were also saved and analyzed. The EE-polypeptide elution peak fractions were analyzed for the target protein via SDS-PAGE Silver staining and Western Blotting with the anti-EE HRP conjugated antibody. The polypeptide elution fractions of interest were pooled and
5 concentrated from 60 ml to 5.0 ml using a 10,000 Dalton molecular weight cutoff membrane spin concentrator (Millipore, Bedford, MA) according to the manufacturer's instructions.

To separate IL-TIF-CEE from other co-purifying proteins, the concentrated polypeptide elution pooled fractions were subjected to a POROS HQ-50
10 (strong anion exchange resin from PerSeptive BioSystems, Framingham, MA) at pH 8.0. A 1.0 x 6.0 cm column was poured and flow packed on a BioCad Sprint. The column was counter ion charged then equilibrated in 20mM TRIS pH 8.0 (Tris (Hydroxymethyl Aminomethane)). The sample was diluted 1:13 (to reduce the ionic strength of PBS) then loaded on the Poros HQ column at 5 ml/minute. The column was
15 washed for 10 CVs with 20mM Tris pH 8.0 then eluted with a 40 CV gradient of 20 mM Tris/ 1 M sodium chloride (NaCl) at 10 ml/minute. 1.5 ml fractions were collected over the entire chromatography and absorbance at 280 and 215 nM were monitored. The elution peak fractions were analyzed via SDS-PAGE Silver staining. Fractions of interest were pooled and concentrated to 1.5-2 ml using a 10,000 Dalton molecular
20 weight cutoff membrane spin concentrator (Millipore, Bedford, MA) according to the manufacturer's instructions.

To separate IL-TIF-CEE polypeptide from free EE peptide and any contaminating co-purifying proteins, the pooled concentrated fractions were subjected to size exclusion chromatography on a 1.5 x 90 cm Sephadex S200 (Pharmacia,
25 Piscataway, NJ) column equilibrated and loaded in PBS at a flow rate of 1.0 ml/min using a BioCad Sprint. 1.5 ml fractions were collected across the entire chromatography and the absorbance at 280 and 215 nM were monitored. The peak fractions were characterized via SDS-PAGE Silver staining, and only the most pure fractions were pooled. This material represented purified IL-TIF-CEE polypeptide.

30 This purified material was finally subjected to a 4 ml ActiClean Etox (Sterogene) column to remove any remaining endotoxins. The sample was passed over

the PBS equilibrated gravity column four times then the column was washed with a single 3 ml volume of PBS, which was pooled with the "cleaned" sample. The material was then 0.2 micron sterile filtered and stored at -80°C until it was aliquoted.

On Western blotted, Coomassie Blue and Silver stained SDS-PAGE
5 gels, the IL-TIF-CEE polypeptide was one major band. The protein concentration of the purified material was performed by BCA analysis (Pierce, Rockford, IL) and the protein was aliquoted, and stored at -80°C according to standard procedures.

To prepare anti-EE Sepharose, a 100 ml bed volume of protein G-
10 Sepharose (Pharmacia, Piscataway, NJ) was washed 3 times with 100 ml of PBS containing 0.02% sodium azide using a 500 ml Nalgene 0.45 micron filter unit. The gel was washed with 6.0 volumes of 200 mM triethanolamine, pH 8.2 (TEA, Sigma, St. Louis, MO), and an equal volume of EE antibody solution containing 900 mg of antibody was added. After an overnight incubation at 4°C, unbound antibody was
15 removed by washing the resin with 5 volumes of 200 mM TEA as described above. The resin was resuspended in 2 volumes of TEA, transferred to a suitable container, and dimethylpimilimidate-2HCl (Pierce, Rockford, IL) dissolved in TEA, was added to a final concentration of 36 mg/ml of protein G-Sepharose gel. The gel was rocked at room temperature for 45 min and the liquid was removed using the filter unit as
20 described above. Nonspecific sites on the gel were then blocked by incubating for 10 min. at room temperature with 5 volumes of 20 mM ethanolamine in 200 mM TEA. The gel was then washed with 5 volumes of PBS containing 0.02% sodium azide and stored in this solution at 4°C.

25

Example 13

In vivo affects of IL-TIF polypeptide

Mice (female, C57Bl, 8 weeks old; Charles River Labs, Kingston, NY) were divided into three groups. An adenovirus expressing an IL-TIF polypeptide (SEQ ID NO:8) was previously made using standard methods. On day 0, parental or IL-TIF
30 adenovirus was administered to the first (n=8) and second (n=8) groups, respectively, via the tail vein, with each mouse receiving a dose of $\sim 1 \times 10^{11}$ particles in ~ 0.1 ml

volume. The third group (n=8) received no treatment. On days 12, mice were weighed and blood was drawn from the mice. Samples were analyzed for complete blood count (CBC) and serum chemistry. Statistically significant elevations in neutrophil and platelet counts were detected in the blood samples from the IL-TIF adenovirus administered group relative to the parental adenovirus treated group. Also, lymphocyte and red blood cell counts were significantly reduced from the IL-TIF adenovirus administered group relative to the parental adenovirus treated group. In addition, the IL-TIF adenovirus treated mice decreased in body weight, while parental adenovirus treated mice gained weight. The SAA and globulin level was increased and glucose level was decreased. The adeno-zcyto18 mice displayed wasting syndrome. In summary, zcyto18 causes acute phase response (APR) that reflects the pro-inflammatory activity of TNF- α , IL-1, and gp130 cytokines.

The result suggested that IL-TIF is a pro-inflammatory factor that is involved in the immune and inflammatory response *in vivo*. The tissue distribution of IL-TIF receptor (zcytor11) indicated that elevated expression of IL-TIF in circulation or specific tissues can lead to certain acute/chronic inflammatory diseases, such as pancreatitis, IBD (Chron's disease, colitis), asthma, ESRD (end stage renal diseases), rheumatoid arthritis, psoriasis, and autoimmune diseases (GVHD, lupus, sepsis).

The results suggested that IL-TIF affects hematopoiesis, i.e., blood cell formation *in vivo*. As such, IL-TIF could have biological activities effecting different blood stem cells, thus resulting increase or decrease of certain differentiated blood cells in a specific lineage. For instance, IL-TIF appears to reduce lymphocytes, which is likely due to inhibition of the committed progenitor cells that give rise to lymphoid cells. IL-TIF also decreases red blood cells. This finding agrees with the inhibitory effects of IL-TIF on the proliferation and/or growth of myeloid stem cells (Example 13), supporting the notion that IL-TIF could play a role in anemia, infection, inflammation, and/or immune diseases by influencing blood cells involved in these process. Antagonists against IL-TIF, such as antibodies or zcytor11 soluble receptors of the present invention, could be used as therapeutic reagents in these diseases.

Moreover, these experiments using IL-TIF adenovirus in mice suggest that IL-TIF over-expression increases the level of neutrophils and platelets *in vivo*. It is

conceivable that there are other factors (such as cytokines and modifier genes) involved in the responses to IL-TIF in the whole animal system. Nevertheless, these data strongly support the involvement of IL-TIF in hematopoiesis. Thus, IL-TIF and its receptors are suitable reagents/targets for the diagnosis and treatment in variety of disorders, such as inflammation, immune disorders, infection, anemia, hematopoietic and other cancers, and the like.

Example 14

Identification of Cells Expressing *zcycor11* Using *in situ* Hybridization

Specific human tissues were isolated and screened for *zcycor11* expression by *in situ* hybridization. Various human tissues prepared, sectioned and subjected to *in situ* hybridization included appendix, brain, cartilage, colon, intestine, kidney, liver, lung, lymph node, lymphoma, ovary, pancreas, placenta, prostate, skin, spleen, and thymus. The tissues were fixed in 10% buffered formalin and blocked in paraffin using standard techniques. Tissues were sectioned at 4 to 8 microns. Tissues were prepared using a standard protocol ("Development of non-isotopic *in situ* hybridization" at The Laboratory of Experimental Pathology (LEP), NIEHS, Research Triangle Park, NC; web address <http://dir.niehs.nih.gov/dirlep/ish.html>). Briefly, tissue sections were deparaffinized with HistoClear (National Diagnostics, Atlanta, GA) and then dehydrated with ethanol. Next they were digested with Proteinase K (50 µg/ml) (Boehringer Diagnostics, Indianapolis, IN) at 37°C for 2 to 7 minutes. This step was followed by acetylation and re-hydration of the tissues.

One *in situ* probe was designed against the human *zcycor11* sequence (nucleotide 234-1105 in SEQ ID NO:1), and isolated from a plasmid containing SEQ ID NO:1 using standard methods. T7 RNA polymerase was used to generate an antisense probe. The probe was labeled with digoxigenin (Boehringer) using an *In Vitro* transcription System (Promega, Madison, WI) as per manufacturer's instruction.

In situ hybridization was performed with a digoxigenin-labeled *zcycor11* probe (above). The probe was added to the slides at a concentration of 1 to 5 pmol/ml for 12 to 16 hours at 60 °C. Slides were subsequently washed in 2XSSC and 0.1XSSC at 55°C. The signals were amplified using tyramide signal amplification (TSA) (TSA,

in situ indirect kit; NEN) and visualized with Vector Red substrate kit (Vector Lab) as per manufacturer's instructions. The slides were then counter-stained with hematoxylin (Vector Laboratories, Burlingame, CA).

Signals were observed in several tissues tested: The lymph node that
5 contains cells in the paracortex of nodules was strongly positive. In lymphoma samples, there is little to no signal in the samples tested. In spleen, positive signals were seen in scattered mononuclear cells at the periphery of follicles were positive. In thymus, positive signals were seen in scattered mononuclear cells in both cortex and medulla was positive. In fetal liver, a strong signal was observed in a mixed population
10 of mononuclear cells in sinusoid spaces. Some circulating mononuclear cells were also positive. Hepatocytes were negative. In the inflamed appendix, mononuclear cells in Peyer's patch and infiltration sites were positive. In intestine, cells in lamina propria and Peyer's patches were strongly positive. Ganglia nerve cells in the muscles were positive. In normal lung, zcytor11 was expressed in alveolar epithelium and
15 mononuclear cells in interstitial tissue and circulation. In the lung carcinoma tissue, a weak signal was observed in carcinoma cells and mononuclear cells in peripheral of carcinoma sites. In ovary carcinoma, epithelium cells were strongly positive. Some interstitial cells, most likely the mononuclear cells, were also positive. There was no signal observed in the normal ovary. In kidney, podocytes and simple epithelial cells in
20 the parietal layer of Bowman's capsules in renal corpuscles were positive. Cuboidal epithelial cells of distal convoluted tubules were also positive. In both normal and pancreatitis pancreas samples, acinar cells and some mononuclear cells in the mesentery were positive. There may be also weak signal in a subset of cells in islets of pancreas. In the early term (8 weeks) placenta, signal was observed in trophoblasts. In skin,
25 strong signal was observed in the keratinocytes and mononuclear cells in the inflamed infiltrates in the superficial dermis. In brain, majority of neurons in temporal lobe were positive, however, the frontal lobe appears to be negative. In articular cartilage, chondrocytes were positive. Other tissues tested including normal ovary, skin melanoma, prostate carcinoma and BPH were negative.

30 In summary, the *in situ* data was consistent with expression data described above for the zcytor11. Zcytor11 expression was observed predominately

and consistently expressed by a mixed population of mononuclear cells. A subset of epithelium was also positive. These results confirmed the presence of zcytor11 expression in immune cells and point toward a role in inflammation, autoimmune disease, or other immune function, for example, in binding pro-inflammatory cytokines, including but not limited to IL-TIF. Moreover, detection of zcytor11 expression can be used for example as a marker for mononuclear cells in histologic samples.

Zcytor11 is expressed in mononuclear cells, including normal tissues (lymph nodes, spleen, thymus, pancreas, kidney, liver and lung), and abnormal tissues (inflamed appendix, lung carcinoma, ovary carcinoma, pancreatitis, inflamed skin, and prostate carcinoma). It is notable that plasma cells in the lymph node, intestine, and lung carcinoma are positive for zcytor11. Plasma cells are immunologically activated lymphocytes responsible for antibody synthesis. In addition, IL-TIF is expressed in activated T cells, and the expression of zcytor11 is detected only in resting (but not in activated) CD19+ cells (Example 13). Thus, zcytor11 can be used as a marker for or as a target in isolating certain lymphocytes, such as mononuclear leucocytes and limited type of activated leucocytes, such as resting CD19+.

Furthermore, the presence of zcytor11 expression in immune cells such as CD8+ T cell and CD19+ B cells showed that zcytor11 may be involved in the body's immune defensive reactions against foreign invaders: such as microorganisms and cell debris, and could play a role in immune responses during inflammation and cancer formation. The activation of zcytor11 receptor may cause autoimmune and inflammatory diseases such as GVHD, sepsis and lupus.

Moreover, as discussed herein, epithelium from several tissues was positive for zcytor11 expression, such as skin, kidney, gut, hepatocytes (endoderm-derived epithelia), lung alveolar epithelium (endoderm-derived epithelia), and ovary carcinoma epithelium (mesoderm-derived epithelium). The inflammatory response in these tissues may cause acute/chronic inflammatory diseases, such as psoriasis (skin), end-stage renal disease ESRD (kidney), IBD (Chron's disease, colitis) (gut), and asthma/respiratory allergy/chronic bronchitis (lung). The epithelium expression of zcytor11 could be altered in inflammatory responses and/or cancerous states in liver and lung. Thus, ligand for zcytor11, such as IL-TIF, or a receptor-binding fragment thereof,

could be used as marker to monitor changes in these tissues as a result of inflammation or cancer. Moreover, analysis of zcytor11 *in situ* expression showed that normal ovary epithelium is negative for zcytor11 expression, while it is strongly positive in ovary carcinoma epithelium providing further evidence that IL-TIF polypeptides, or a
 5 receptor-binding fragment thereof, can be used as a diagnostic marker and/or therapeutic target for the diagnosis and treatment of ovarian cancers, and ovary carcinoma, as described herein.

Zcytor11 was also detected in other tissues, such as acinar cells in pancreas (normal and pancreatitis tissues), trophoblasts in placenta (ectoderm-derived),
 10 chondrocytes in cartilage (mesoderm-derived), and ganglia cells in intestine (ectoderm-derived). As such, zcytor11 may be involved in differentiation and/or normal functions of corresponding cells in these organs. As such, potential utilities of zcytor11 include maintenance of normal metabolism and pregnancy, bone formation/homeostasis, and physiological function of intestine, and the like. Moreover, the up-regulation of IL-TIF
 15 may potentially cause the inflammatory response in those tissues that lead to certain inflammatory diseases such as pancreatitis, rheumatoid arthritis, IBD (colitis and Chron's disease).

Example 15

20 Human zcytor11 Tissue Distribution in Tissue Panels Using Northern Blot and PCR A. Human zcytor11 Tissue Distribution in tissue panels using PCR

A panel of cDNAs from human tissues was screened for zcytor11 expression using PCR. The panel was made in-house and contained 94 marathon cDNA and cDNA samples from various normal and cancerous human tissues and cell lines are
 25 shown in Table 4 below. Aside from the PCR reaction, the method used was as shown in Example 12. The PCR reactions were set up using oligos ZC14,666 (SEQ ID NO: 11) and ZC14,742 (SEQ ID NO:12), Advantage 2 cDNA polymerase mix (Clontech, Palo Alto, CA), and Rediload dye (Research Genetics, Inc., Huntsville, AL). The amplification was carried out as follows: 1 cycle at 94°C for 2 minutes, 40 cycles of
 30 94°C for 15 seconds, 51°C for 30 seconds and 72°C for 30 seconds, followed by 1 cycle at 72°C for 7 minutes. The correct predicted DNA fragment size was observed in

bladder, brain, cervix, colon, fetal brain, fetal heart, fetal kidney, fetal liver, fetal lung, fetal skin, heart, kidney, liver, lung, melanoma, ovary, pancreas, placenta, prostate, rectum, salivary gland, small intestine, testis, thymus, trachea, spinal cord, thyroid, lung tumor, ovarian tumor, rectal tumor, and stomach tumor. Zcytor11 expression was not
 5 observed in the other tissues and cell lines tested in this panel.

A commercial 1st strand cDNA panel (Human Blood Fractions MTC Panel, Clontech, Palo Alto, CA) was also assayed as above. The panel contained the following samples: mononuclear cells, activated mononuclear cells, resting CD4+ cells, activated CD4+ cells, resting CD8+ cells, activated CD8+ cells, resting CD14+
 10 cells, resting CD19+ cells and activated CD19+ cells. All samples except activated CD8+ and Activated CD19+ showed expression of zcytor11.

Table 4

<u>Tissue</u>	<u>#samples</u>		<u>Tissue</u>	<u>#samples</u>
adrenal gland	1		bladder	1
bone marrow	3		brain	2
cervix	1		colon	1
fetal brain	3		fetal heart	2
fetal kidney	1		fetal liver	2
fetal lung	1		fetal skin	1
heart	2		fetal muscle	1
kidney	2		liver	1
lung	1		lymph node	1
mammary gland	1		melanoma	1
ovary	1		pancreas	1
pituitary	2		placenta	3
prostate	3		rectum	1
salivary gland	2		skeletal muscle	1
small intestine	1		spinal cord	2
spleen	1		uterus	1
stomach	1		adipocyte library	1
testis	5		islet	1
thymus	1		prostate SMC	1
thyroid	2		RPMI 1788	1
trachea	1		WI38	1
esophageal tumor	1		lung tumor	1
liver tumor	1		ovarian tumor	1
rectal tumor	1		stomach tumor	1
uterine tumor	2		CD3+ library	1

HaCAT library	1		HPV library	1
HPVS library	1		MG63 library	1
K562	1			

B. Tissue Distribution of Zcytor11 in Human Cell Line and Tissue Panels Using RT-PCR

A panel of RNAs from human cell lines was screened for zcytor11 expression using RT-PCR. The panels were made in house and contained 84 RNAs from various normal and cancerous human tissues and cell lines as shown in Tables 5-8 below. The RNAs were made from in house or purchased tissues and cell lines using the RNAeasy Midi or Mini Kit (Qiagen, Valencia, CA). The panel was set up in a 96-well format with 100 ngs of RNA per sample. The RT PCR reactions were set up using oligos ZC14,666 (SEQ ID NO:11) and ZC14,742 (SEQ ID NO:12), Rediload dye and SUPERSCRIPT One Step RT-PCR System(Life Technologies, Gaithersburg, MD). The amplification was carried out as follows: one cycle at 50° for 30 minutes followed by 45 cycles of 94°, 15 seconds; 52°, 30 seconds; 72°, 30 seconds; then ended with a final extension at 72° for 7 minutes. 8 to 10 uls of the PCR reaction product was subjected to standard Agarose gel electrophoresis using a 4% agarose gel. The correct predicted cDNA fragment size was observed in adrenal gland, bladder, breast, bronchus, normal colon, colon cancer, duodenum, endometrium, esophagus, gastic cancer, gastro-esophageal cancer, heart ventricle, iluem, normal kidney, kidney cancer, liver, lung, lymph node, pancreas, parotid, skin, small bowel, stomach, thyroid, and uterus. Cell lines showing expression of zcytor11 were A-431, differentiated CaCO2, DLD-1, HBL-100, HCT-15, HepG2, HepG2+IL6, HuH7, and NHEK #1-4. Zcytor11 expression was not observed in the other tissues and cell lines tested in this panel. The expression pattern of zcytor11 shows expression in specific tissues and tissue-specific tumors. One of skill in the art would recognize that the polynucleotides, polypeptides, antibodies, and binding partners of the present invention can be used as a diagnostic to detect cancer, or cancer tissue in a biopsy, tissue, or histologic sample, particularly in tissues where zcytor11 is expressed. Such diagnostic uses for the molecules of the present invention are known in the art and described herein.

In addition, because the expression pattern of zcytor11, one of IL-TIF's receptors, shows expression in certain specific tissues, binding partners including the

natural ligand, IL-TIF, can also be used as a diagnostic to detect specific tissues (normal or abnormal), cancer, or cancer tissue in a biopsy, tissue, or histologic sample, particularly in tissues where IL-TIF receptors are expressed. IL-TIF can also be used to target other tissues wherein its receptors, e.g., zcytor11 and CRF2-4, are expressed.

- 5 Moreover, such binding partners could be conjugated to chemotherapeutic agents, toxic moieties and the like to target therapy to the site of a tumor or diseased tissue. Such diagnostic and targeted therapy uses are known in the art and described herein.

The expression patterns of zcytor11 (above) indicated target tissues and cell types for the action of IL-TIF, and hence IL-TIF antagonists, such as the soluble
10 zcytor11 receptors of the present invention. The zcytor11 is generally expressed in three physiologic systems: digestive system, female reproductive system, and immune system. Moreover, the expression pattern of the receptor (zcytor11) indicated that an IL-TIF antagonist the soluble zcytor11 receptors of the present invention would have therapeutic application for human disease in two areas: inflammation (e.g., IBD,
15 Chron's disease, pancreatitis) and cancer (e.g., ovary, colon). That is, the polynucleotides, polypeptides and antibodies of the present invention can be used to antagonize the inflammatory, and other cytokine-induced effects of IL-TIF interaction with the cells expressing the zcytor11 receptor.

Moreover, the expression of zcytor11 appeared to be downregulated or
20 absent in an ulcerative colitis tissue, HepG2 liver cell line induced by IL-6, activated CD8+ T-cells and CD19+ B-cells. These RT-PCR experiments demonstrate that CD19+ peripheral blood cells, B lymphocytes, express receptors for IL-TIF, namely zcytoR11. The soluble zcytor11 receptors of the present invention would act as an antagonist to neutralize the effects of IL-TIF on B cells. This would be beneficial in
25 diseases where B cells are the key players: Autoimmune diseases including systemic lupus erythmatosus (SLE), myasthenia gravis, immune complex disease, and B-cell cancers that are exacerbated by IL-TIF. Also autoimmune diseases where B cells contribute to the disease pathology would be targets for zcytor11 soluble receptor therapy: Multiple sclerosis, inflammatory bowel disease (IBD) and rheumatoid arthritis
30 are examples. Soluble zcytor11 receptor therapy would be beneficial to dampen or

inhibit B cells producing IgE in atopic diseases including asthma, allergy and atopic dermatitis where the production of IgE contributes to the pathogenesis of disease.

B cell malignancies may exhibit a loss of regulation by cytokines, such as IL-TIF. The administration of the soluble zcytoR11 receptors of the present invention
5 following surgical resection or chemotherapy may be useful to treat minimal residual disease in patients with B cell malignancies. The loss of regulation may lead to sustain or increased expression of zcytoR11. Thus creating a target for therapeutic monoclonal antibodies targeting zcytoR11 comprising receptors, such as those described herein.

Table 5

Tissue	#samples	Tissue	#samples
adrenal gland	6	duodenum	1
bladder	3	endometrium	5
brain	2	cancerous endometrium	1
brain meningioma	1	gastric cancer	1
breast	1	esophagus	7
cancerous breast	4	gastro-esophageal	1
normal breast adjacent to cancer	5	heart aorta	1
bronchus	3	heart left ventricle	4
colon	15	heart right ventricle	2
cancerous colon	1	heart ventricle	1
normal colon adjacent to cancer	1	ileum	3
ulcerative colitis colon	1	kidney	15
		cancerous kidney	1

Table 6

Tissue/Cell Line	#samples	Tissue/Cell Line	#samples
293	1	HBL-100	1
C32	1	Hs-294T	1
HaCat#1	1	Molt4	1
HaCat#2	1	RPMI	1
HaCat#3	1	U-937	1
HaCat#4	1	A-375	1
WI-38	1	HCT-15	1
WI-38 + 2 um ionomycin #1	1	HT-29	1
WI-38 + 2 um ionomycin #2	1	MRC-5	1
WI-38 + 5 um ionomycin#1	1	RPT-1	1
WI-38 + 5 um ionomycin#2	1	RPT-2	1
Caco-2,	1	WM-115	1
Caco-2,differentiated	1	A-431	1
DLD-1	1	WERI-Rb-1	1
HRE	1	HEL-92.1.7	1
HRCE	1	HuH-7	1
MCF7	1	MV-4-11	1
PC-3	1	U-138	1
TF-1	1	CCRF-CEM	1

5637	1	Y-79	1
I43B	1	A-549	1
ME-180	1	EL-4	1
prostate epithelia	1	HeLa 229	1
U-2 OS	1	HUT 78	1
T-47D	1	NCI-H69	1
Mg-63	1	SaOS2	1
Raji	1	USMC	1
U-373 MG	1	UASMC	2
A-172	1	AoSMC	1
CRL-1964	1	UtSMC	1
CRL-1964 + butyric acid	1	HepG2	1
HUVEC	1	HepG2- IL6	1
SK-Hep-1	1	NHEK#1	1
SK-Lu-1	1	NHEK#2	1
Sk-MEL-2	1	NHEK#3	1
K562	1	NHEK#4	1
BeWo	1	ARPE-19	1
FHS74.Int	1	G-361	1
HL-60	1	HISM	1
Malme 3M	1	3AsubE	1
FHC	1	INT407	1
HREC	1		

Table 7

Tissue	#samples	Tissue	#samples
liver	10	lung	13
lymph node	1	cancerous lung	2
lymphoma	4	normal lung adjacent to cancer	1
mammary adenoma	1	muscle	3
mammary gland	3	neuroblastoma	1
melinorioma	1	omentum	2
osteogenic sarcoma	2	ovary	6
pancreas	4	cancerous ovary	2
skin	5	parotid	7
sarcoma	2	salivary gland	4

Table 8

Tissue	#samples	Tissue	#samples
small bowel	10	uterus	11
spleen	3	uterine cancer	1

spleen lymphoma	1	thyroid	9
stomach	13		
stomach cancer	1		

C. Tissue Distribution of Zcytor11 in Human Origene™ Tissue and Human Blood Fractions MTC Panels Using RT-PCR

A panel of RNAs from human tissues, Human Origene™ Tissue and Human Blood Fractions MTC Panels (Origene Technologies, Rockville, MD; and Clontech, Palo Alto, CA) was screened for zcytor11 expression using RT-PCR. The panels contained 24 RNAs from various normal human tissues at increasing concentrations as shown in Tables 9-10 below. The RT-PCR reactions were set up using oligos ZC37693 (SEQ ID NO:31) and ZC37449 (SEQ ID NO:32), using the Advantage PCR kit (Clontech). The amplification was carried out as follows: one cycle at 94°C for 2 min.; 35 cycles of 94°C for 15 sec., 72°C for 1.5 min; then 72°C for 2 min.; followed by a 4°C hold. 8 to 10 µl of the PCR reaction product was subjected to standard Agarose gel electrophoresis using a 4% agarose gel.

Using the Origene Panel, as shown in Table 9, the correct predicted cDNA fragment size (440 bp) was observed in all tissues except spleen, muscle, placenta, PBL, bone marrow and fetal brain. However, there was high expression specifically in small intestine, colon, kidney, skin, lung, pancreas and liver. The weak expression of zcytor11 was also observed in ovary, uterus, prostate, brain, heart, testis, stomach, and thyroid. Zcytor11 expression was not observed in the other tissues and cell lines tested in this panel. The expression pattern of zcytor11 shows expression in specific tissues. One of skill in the art would recognize that the polynucleotides, polypeptides, antibodies, and binding partners of the present invention can be used as a diagnostic to detect such tissues, cancer, or cancer tissue in a biopsy, tissue, or histologic sample, particularly in tissues where zcytor11 is expressed. Such diagnostic uses for the molecules of the present invention are known in the art and described herein.

Using the Human Blood Fractions MTC Panel, as shown in Table 10, the correct predicted cDNA fragment size (440 bp) was observed in mononuclear cells, resting CD8+ cells, resting CD19+ cells and placenta. Zcytor11 expression was not

observed in the other tissues and cell lines tested in this panel, including activated CD8+ and CD19+ cells. The expression pattern of zcytor11 shows expression in specific tissues. One of skill in the art would recognize that the polynucleotides, polypeptides, antibodies, and binding partners of the present invention can be used as a

5 diagnostic to detect such tissues, cancer, or cancer tissue in a biopsy, tissue, or histologic sample, particularly in tissues where zcytor11 is expressed. Such diagnostic uses for the molecules of the present invention are known in the art and described herein.

10

Table 9

Tissue	1pg cDNA	10pg cDNA	100pg cDNA	1ng cDNA
Brain				X
Heart				X
Kidney			X	X
Spleen				
Liver			X	X
Colon			X	X
Lung			X	X
Small Intestine		X	X	X
Muscle				
Stomach			X	X
Testis	X		X	X
Placenta			X	
Salivary Gland				X
Thyroid Gland			X	X
Adrenal Gland			X	X
Pancreas	X	X	X	X
Ovary				X
Uterus				X
Prostate				X
Skin			X	X
PBL				
Bone Marrow				
Fetal Brain				
Fetal Liver		X	X	X

Table 10

1ng cDNA	Expression
Mononuclear cells	X
Resting CD8+ cells	X
Resting CD4+ cells	
Resting CD14+ cells	
Resting CD19+cells	X
Activated CD19+cells	
Activated mononuclear cells	
Activated CD4+ cells	
Activated CD8+ cells	
Human Placenta	X
No template control	
Zcytor11 cDNA	X

D. Tissue Distribution of Zcytor11 in Human Primary Immune Cell and Immune Cell

5 Lines Using RT-PCR

A panel of RNAs from primary human immune cell populations and human immune cell lines was screened for zcytor11 expression using RT-PCR. The panels were made in house and contained 24 RNAs from various resting and activated cell populations and cell lines as shown in Table 11 below. All primary immune cell

10 populations were isolated from the blood of several anonymous donors. Various immune cell subsets (CD4+, CD8+, CD14+, CD19+, and CD56+) were then isolated using Microbeads and the Magnetic Cell Separation System from Miltenyi Biotec. RNA was prepared from the CD19+ and CD56+ populations in their resting state using an RNeasy Midiprep™ Kit (Qiagen, Valencia, CA) as per manufacturer's instruction.

15 The CD4+, and CD8+ populations were activated using 200 ng/ml plate-bound anti-CD3 antibody and 5 µg/ml soluble anti-CD28 antibody and cells were collected for RNA isolation at 0, 4 and 16 hours. The CD19+ samples were isolated from human tonsil and activated with 0.5 µg/ml Ionomycin and 10ng/ml PMA. Cells were then collected at 0, 4 hours and 24 hours and RNA isolated. Human CD14+ monocytes

20 were activated with either 0.1µg/ml LPS or 1.0 µg/ml LPS for 20 hours. Resting and activated cells were then collected and RNA isolated. In addition, RNA was isolated

from resting and activated (10.0 µg/ml LPS) human monocyte cell lines HL-60, THP-1 and U937. Also, resting Raji, Ramos, Daudi, and Jurkat RNA's were tested.

The RT-PCR reactions used the Superscript One-Step RT-PCR System with Platinum Taq. Each 25 µl reaction consisted of the following: 12.5 µl of 2X
 5 Reaction Buffer, 0.5 µl (20pmol/µl) ZC14,666 (SEQ ID NO:11), 0.5 µl (20pmol/µl) ZC14,742 (SEQ ID NO:12), 0.4 µl RT/Taq polymerase mix, 10 µl RNase-free water, 1.0 µl template RNA (100ng/µl). (Life Technologies, Gaithersburg, MD). The amplification was carried out as follows: one cycle at 50° for 30 minutes followed by 35 cycles of 94°, 30 seconds; 52°, 30 seconds; 72°, 60 seconds; then ended with a final
 10 extension at 72° for 7 minutes. 8 to 10 µl of the PCR reaction product was subjected to standard Agarose gel electrophoresis using a 2% agarose gel. The correct predicted cDNA fragment size was observed in resting CD19+ B cells and much less in the activated CD19+ B cells, resting CD8+ T cells, CD56+ NK cells and activated CD14+ monocytes. Cell lines showing expression of zcytor11 were Jurkat, activated THP-1 and
 15 activated HL-60. Zcytor11 expression was not observed in the other tissues and cell lines tested in this panel. These results demonstrate the expression of zcytor11 in several immune cell populations and immune cell lines.

Table 11.

Tissue	#samples		Tissue	#samples
CD19+ from resting PBMCs	1		CD14+ 0.1 µg/ml LPS	1
CD19+ resting from tonsil	1		CD14+ 1.0 µg/ml LPS	1
CD19+ from tonsil -- 4hr activation	1		Raji	1
CD19+ from tonsil -- 16hr activation	1		Ramos	1
CD4+ Resting	1		Daudi	1
CD4+ 4hr Activation	1		Jurkat	1
CD4+ 16hr Activation	1		U937	1
CD8+ Resting	1		Activated U937	1
CD8+ 4hr Activation	1		THP-1	1
CD8+ 16hr Activation	1		Activated THP-1	1
CD56+ Resting	1		HL-60	1
CD14+ Resting	1		Activated HL-60	1

Example 16

Construction of BaF3 Cells Expressing the CRF2-4 Receptor (BaF3/CRF2-4 cells) and BaF3 Cells Expressing the CRF2-4 Receptor With the zcytor11 Receptor (BaF3/CRF2-4/zcytor11 cells)

5 BaF3 cells expressing the full-length CFR2-4 receptor were constructed, using 30µg of a CFR2-4 expression vector, described below. The BaF3 cells expressing the CFR2-4 receptor were designated as BaF3/CFR2-4. These cells were used as a control, and were further transfected with full-length zcytor11 receptor (US Patent No. 5,965,704) and used to construct a screen for IL-TIF activity as described
10 below.

A. Construction of BaF3 Cells Expressing the CRF2-4 receptor

The full-length cDNA sequence of CRF2-4 (Genbank Accession No. Z17227) was isolated from a Daudi cell line cDNA library, and then cloned into an
15 expression vector pZP7P.

BaF3, an interleukin-3 (IL-3) dependent pre-lymphoid cell line derived from murine bone marrow (Palacios and Steinmetz, Cell 41: 727-734, 1985; Mathey-Prevot et al., Mol. Cell. Biol. 6: 4133-4135, 1986), was maintained in complete media (RPMI medium (JRH Bioscience Inc., Lenexa, KS) supplemented with 10% heat-
20 inactivated fetal calf serum, 2 ng/ml murine IL-3 (mIL-3) (R & D, Minneapolis, MN), 2 mM L-glutaMax-1™ (Gibco BRL), 1 mM Sodium Pyruvate (Gibco BRL), and PSN antibiotics (GIBCO BRL)). Prior to electroporation, CRF2-4/pZP7P was prepared and purified using a Qiagen Maxi Prep kit (Qiagen) as per manufacturer's instructions. For electroporation, BaF3 cells were washed once in serum-free RPMI media and then
25 resuspended in serum-free RPMI media at a cell density of 10^7 cells/ml. One ml of resuspended BaF3 cells was mixed with 30 µg of the CRF2-4/pZP7P plasmid DNA and transferred to separate disposable electroporation chambers (GIBCO BRL). Following a 15-minute incubation at room temperature the cells were given two serial shocks (800 IFad/300 V.; 1180 IFad/300 V.) delivered by an electroporation apparatus (CELL-
30 PORATOR™; GIBCO BRL). After a 5-minute recovery time, the electroporated cells were transferred to 50 ml of complete media and placed in an incubator for 15-24 hours

(37°C, 5% CO₂). The cells were then spun down and resuspended in 50 ml of complete media containing 2 µg/ml puromycin in a T-162 flask to isolate the puromycin-resistant pool. Pools of the transfected BaF3 cells, hereinafter called BaF3/CRF2-4 cells, were assayed for signaling capability as described below. Moreover these cells were further transfected with zcytor11 receptor as described below.

B. Construction of BaF3 Cells Expressing CRF2-4 and zcytor11 receptors

BaF3/CRF2-4 cells expressing the full-length zcytor11 receptor were constructed as per Example 5A above, using 30µg of the zcytor11 expression vector, described in Example 6 above. Following recovery, transfectants were selected using 200µg/ml zeocin and 2µg/ml puromycin. The BaF3/CRF2-4 cells expressing the zcytor11 receptor were designated as BaF3/CRF2-4/zcytor11 cells. These cells were used to screen for IL-TIF activity as well as zcytor16 antagonist activity described IN Example 17.

Example 17

Screening for CRF2-4Zcytor11-Fc Activity Using BaF3/CRF2-4/zcytor11 Cells in an Alamar Blue Proliferation Assay

BaF3/CRF2-4/zcytor11 cells (Example 16) were spun down and washed in PBS 2 times to ensure the removal of the mIL-3, and then spun a third time and re-suspended in the complete media, (RPMI 1640, 10%FBS, 1% GlutaMAX, 1% Sodium Pyruvate) described in Example 16 above, but without mIL-3 (hereinafter referred to as "mIL-3 free media"). Cells were then counted in a hemocytometer. Cells were plated in a 96-well format at 5000 cells per well in a volume of 50 µl per well using the mIL-3 free media.

IL-TIF protein was diluted to 200 pg/ml in mIL-3 free media and also in CRF2-4/zcytor11-Fc conditioned media at a concentration of approximately 0.4 ug/ml that was made from transfected BHK cells (Example 7). The CRF2-4/zcytor11-Fc CM was diluted into the mIL-3 free/IL-TIF media by serial 1:2 dilutions down all 8 rows on the 96-well plate, leaving a volume of 50 ul in each well. This was then added to the 50

ul of cells, for a final IL-TIF concentration of 100 pg/ml in all wells, and final CRF2-4/zcytor11-Fc concentrations of approximately 200, 100, 50, 25, 12.5, 6.25, 3.1 and 1.6 ng/ml, and a total assay volume of 100 μ l. The assay plates were incubated at 37°C, 5% CO₂ for 4 days at which time Alamar Blue (Accumed, Chicago, IL) was added at 20 μ l/well. Plates were again incubated at 37°C, 5% CO₂ for 24 hours. Alamar Blue gives a fluourometric readout based on number of live cells, and is thus a direct measurement of cell proliferation in comparison to a negative control. Plates were read on the Wallac Victor 2 1420 Multilabel Counter (Wallac, Turku, Finland) at wavelengths 530 (Excitation) and 590 (Emmssion).

Results confirmed a dose-dependant inhibition by CRF2-4/zcytor11-Fc of the proliferative effect of IL-TIF on BaF3/CRF2-4/zcytor11 cells. IL-TIF alone stimulated the cells 30 -fold over background. CRF2-4/zcytor11-Fc completely inhibited that proliferation at concentrations of 25 – 200 ng/ml, partially inhibited proliferation at 3.1 – 12.5 ng/ml, and inhibition was barely detectable at 1.6 ng/ml. The same setup was done with mIL-TIF, and generated similar results.

Example 18

Flow Cytometry Analysis of Baf3-Transfectants Expressing zcytor11 Alone CRF2-4 Alone, zcytor11/CRF-4, or zcytor11/pDIRS1

BaF3 cells transfected with zcytor11 alone, CRF2-4 alone, zcytor11/CRF2-4, or zcytor11/pDIRS1 were generated with each respective cytokine receptor, as described (Example 16). Briefly, 30ug of zcytor11/pZP7Z were transfected into BaF3 cells using electroporation, and the stable transfectants (BaF3/zcytor11) were selected with 200 μ g/ml zeocin. Similarly BaF3 cells transfected with CRF2-4/pZP7P (BaF3/CRF2-4) were selected with 2 μ g/ml puromycin. Subsequently, 30ug of CRF2-4/pZP7P were transfected into BaF3/zcytor11 cells using electroporation, and the stable cell line (BaF3/zcytor11/CRF2-4) was selected with 200 μ g/ml zeocin and 2 μ g/ml puromycin. Similarly, 30ug of pDIRS1/pZP7P were transfected into BaF3/zcytor11 cells using electroporation, and the stable cell line (BaF3/zcytor11/pDIRS1) was selected with 200 μ g/ml zeocin and 2 ug/ml puromycin. The BHK transfectants were generated using the same expression vectors of either zcytor11, CRF2-4, or pDIRS1 for

BaF3 transfectants. DNA was transfected into BHK cells using Lipofectamine™ (Gibco BRL, Gaithersburg, MD) as per manufacturer's instructions, the selection was started 48-hr post-transfection following the same protocol for BaF3 selection.

5 The biotinylation of human IL-TIF-CEE (Example 12) protein is done as follows: 1.6 ul 10% Tween20, 50ul 1M boric acid (pH8.5), and 42ul of 0.9mg/ml EZ-link Sulfo-NHS-LC-biotin (Pierce, Rockford, IL) dissolved in DMSO were added into 100ul of 2.2mg/ml IL-TIF-CEE. After 1 hr incubation at room temperature, the reaction was quenched with 10ul of 2M glycine for 10 minutes.

10 To test the binding properties of the IL-TIF ligand to several potential receptor components, BaF3 and BHK cells were transfected with expression plasmids including zcytoR11 alone, CRF2-4 alone (Genbank Accession No. Z17227), zcytoR11 (SEQ ID NO:1) and CRF2-4, or zcytoR11 and pDIRS1 (WIPO Publication WO99/46379, Schering Corporation, 1999), as described above. Untransfected BaF3 and BHK cells were included as controls. Cells were resuspended in FACS wash buffer
15 (WB: PBS/1%BSA, supplemented with 3% human Ultraserum (Gemini BioProducts, Calabasas, CA)) counted and 1×10^6 of each type were aliquoted into 5ml polystyrene tubes. Cells were washed and pelleted, then incubated for 20 min on ice with 100 μ l of WB only, or WB plus 10 μ g/ml or 1 μ g/ml biotinylated zcyto10 protein. Cells were washed with 1.5 ml WB and pelleted, then incubated in 100 μ l of 2.5 μ g/ml
20 phycoerthyrin-conjugated streptavidin (PE-SA, PharMingen, San Diego, CA) for another 20 min on ice. Cells were washed as before, resuspended in 0.4 ml of WB and analyzed on a FACScan using CellQuest software (Becton Dickinson, Mountain View, CA).

The biotin-IL-TIF bound in a dose-dependent fashion to all 3 of the
25 transfected BaF3 cell lines containing zcytor11 (i.e. zcytor11 transfected alone, or in combination with CRF2-4 or DIRS1), but not to the parental line or to the BaF3 x CRF2-4 transfectant. The same results were obtained with the corresponding BHK transfectants.

From the foregoing, it will be appreciated that, although specific
30 embodiments of the invention have been described herein for purposes of illustration,

various modifications may be made without deviating from the spirit and scope of the invention. Accordingly, the invention is not limited except as by the appended claims.